

Structural and Beneficial Aspects of Tea (*Camellia sinensis*) Flowers

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RUNNING TITLE: Structure and Benefits of Tea Flower

Abstract

Tea is an economical plant for North East India and some parts of South India. Flower morphology is important to plant breeding as they provide important information on flower nature. There are three variety of tea races found in India. China variety i.e. *Camellia sinensis*, Assam variety i.e. *Camellia asamica*, Cambod variety *Camellia asamica sub. lasiocalyx*. Three varieties have different flower morphology by which variety could be identified. Flowering normally starts in monsoon and fruiting starts in early winter.

Keywords

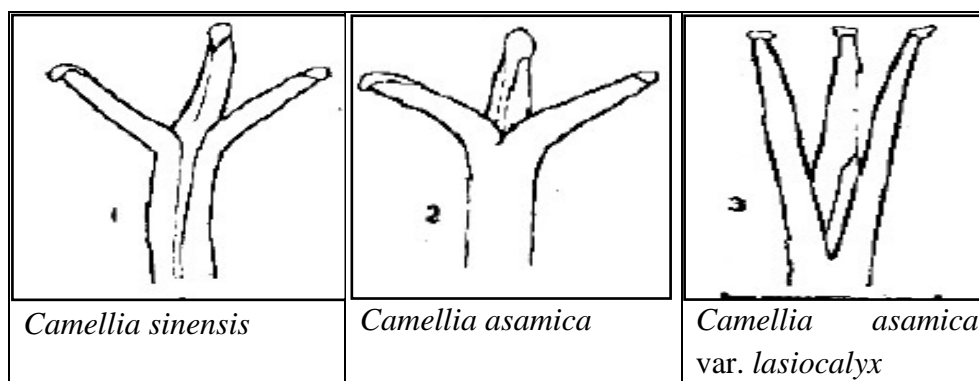
Actinomorphic, Anthesis, Corolla, Gibberellic acid, Pollination, Pserphid, Tea

Introduction

Floral morphology

Tea flowers are actinomorphic i.e. they can be divided into single plane. Wu was the first one to describe morphology of tea flowers. He reported variation between china variety flowers. According to him, classification should be done on the basis of central placenta not by parietal placenta. There are two whorls in the calyx and there are two to four whorls in corolla (Ramasubramanian, 2005).

Tea also can be classified on the basis of the styles. In an study it was revealed that *Camellia sinensis* i.e. China variety styles are more united than others. In *Camellia asamica* i.e. in Assam variety styles are less united than *sinensis* but more than its sub species *lasiocalyx* i.e. cambod. Finally *Camellia asamica sub. lasiocalyx* normally exhibits least united style structure (Curr, 1962).



Pollination

Tea can exhibit both self and cross fertilization. Pollen grains may be triangular to spherical. In a research result showed a flower can have near about 80% viable pollen (Tsou, 1997; Iqbal and Wijesikare, 2002). Pollination occurs through small insects mostly by 'psephid'. These insects have a short flying range. Through keeping distance between two flowering plants chances of cross pollination can be lowered.

Flower buds are visible throughout the year. Flowering normally occurs after July/August. But it can vary depending on agroclimate zone. Seed set up starts after September. Sometimes two three flowering peaks also can be observed in a season. Mature seeds can be observed collected from November to February. Anthesis occurs normally during the morning sunshine from 7.30 to 9.00 A.M. Petal unfurling can continue till noon (Ariyaratna *et al.*, 2011).

Biological functions of Tea Flower

Tea flowers possess many functional metabolites related to polysaccharides, catechins and saponins. Catechins and polysaccharides are responsible for the antioxidant abilities of tea flower. Tea flower polysaccharides have equivalent amount of polysaccharides as tea leaves.

Li *et al.* reported that tea flowers have acute and subchronic toxic effects on rats (Li *et al.*, 2011). Crude tea polysaccharides

Genetics of tea flowering

The process of flower induction and development involves an intricate physiological process comprising of numerous endogenous as well as extraneous factors (Liu *et al.*, 2017). During the past decade, molecular and genetic mechanisms involved in tea flower induction, differentiation and development have been inferred. Floral induction has been found to be influenced by the expression of genes such as gibberellic acid intensive dwarf 1B (GID1B) and GID1C, gibberellin 3-oxidase 1 (GA3ox1), GIGANTEA (GI), pseudo-response regulator (PRR7) and flowering locus T (FT), whereas expression of genes such as leafy (LFY), pound-floolish (PNF) and pennywise (PNY) were correlated with floral bud formation (Liu *et al.*, 2020). 207 unigenes and transcription factors such as WRKY, ERF, MYB, bHLH and

MADS-box have been particularly identified with flowering-associated roles in tea (Liu *et al*, 2017).

Tea flower differentiation and development includes various processes. The tubulin-encoding *Tual* and pollen coat protein (*Pcp*) genes have been observed to promote pollen tube growth (Fang *et al*, 2006) and anther development (Ye *et al*, 2008) in tea. Pollen tube elongation in tea flowers is also regulated via the nitric oxide (NO) pathway under low temperature stress by the CAMTA TFs, COBRA-like genes and phosphatidylinositol-4-kinase (PI4K) (Pan *et al*, 2016).

Moreover, major genes responsible for total catechin content in tea flowers such as chalcone synthase (CH2) and flavonol synthase (FLS) were found to be highly expressed during early flowering stage, while genes such as phenylalanine ammonia lyase (PAL1) and flavonoid 3'-hydroxylase (F3'H1) were expressed in the late flowering stage and negatively correlated with the total catechins content in the flowers (Sun *et al*, 2019). As is evident, aroma compound formation during tea flowering increases. This occurs due to the increasing activity of hydrolytic enzymes such as glycosidases (Watanabe *et al*, 1993; Hayashi *et al*, 2004). Consequently, flavor precursors formed during anthesis are converted to volatile compounds leading to the gradual development of unique odors (Watanabe *et al*, 1993).

Bioactivities

Antioxidant

Tea flowers have a considerable amount of catechins, comparable to the leaves (Lin *et al*, 2003), which in turn contributes to the antioxidant activity of tea flower extract. Ethanol extracts of tea flowers were found to exhibit high scavenging activity against hydroxyl radical ($IC_{50} = 19.7 \mu\text{g/mL}$) and DPPH radical ($IC_{50} = 47.6 \mu\text{g/mL}$) (Yang *et al*, 2007). Isolation and purification of catechins showed the high concentration of EGCG and ECG in tea flowers contributing immensely to its antioxidant activity (Yang *et al*, 2009). The method used for extraction of bioactive compounds from flowers also influence their antioxidant efficiency. Flower extract obtained through super critical extraction (SFE) method was found to retain volatiles (Xia *et al*, 2018) which would otherwise have been degraded due to higher temperatures used for extraction in other processes (Shi *et al*, 2018). In addition to catechins, tea flower polysaccharides also contribute to its antioxidant property (Wang *et al*, 2012).

Anticancer

The anti-tumour and anti-cancerous activity of tea flowers have been recorded extensively in the last decade. Inhibitory effect on growth of transplanted S180 in addition to prolonged mice survival, promotion of plasma interleukin-2, interferon- γ levels and improvement of the T-lymphocyte subsets CD4+ and CD4+/CD8+ percentages was observed after 10 day continued administration of tea flower polysaccharides (TFPS). Furthermore, TFPS also significantly enhanced delayed-type hypersensitivity response and macrophage phagocytosis (Han *et al*, 2010). The water extracts of tea flowers from six different species of *Camellia* – *Camellia japonica*, *Camellia tenuifolia*, *Camellia oleifera*, 2 savoury *Camellias* and *Camellia sinensis*, were used to test the anti-proliferative and

apoptotic effects in human breast cancer MCF-7 cells. The water extract of *Camellia sinensis* was found to be the most active among all the test species, which may be attributed to the presence of (-)-epigallocatechin-3-gallate and (-)-epigallocatechin in *Camellia sinensis* which were absent in other species (Way *et al*, 2009). In another study (Xu *et al*, 2012), various fractions of hot water extracts of tea flowers containing crude fractions of tea flower polysaccharides (TFPS) were found to possess inhibitory activity on the growth of human gastric cancer BGC-823 cells. Wang *et al* (2017) evaluated the anticancerous properties of tea flower saponins (TFS) using human ovarian cancer cell lines. They demonstrated that TFS produced significant anti-proliferative effects against A2780/CP70 and OVCAR-3 cells by inducing p53-dependent apoptosis and S phase arrest via inhibition of the expression of Cdc25A, Cdk2, and CyclinD1 and upregulation of Cyclin E and Cyclin A.

Antiinflammatory

Tea flowers were found to possess adequate amount of anti-inflammatory properties. Chen *et al* (2012) found that administration of tea flower extract effectively inhibited lipopolysaccharide-induced liver inflammation in mice. Furthermore, the concentration of nitric oxide (NO), tumour necrosis factor- α (TNF- α) and interleukin-1 β (IL-1 β) mRNA were found to markedly reduce in mice with immunological liver inflammation after treatment with tea flower extract.

Antiobesity

Due to the adoption of urban lifestyle by most people, obesity has become an ever increasing issue of concern in the present age. Tea has been well known for ages to counteract obesity (Westerterp-Plantenga, 2010; Rains *et al*, 2011; Heber *et al*, 2014). In recent times, a methanolic extract of tea flowers has also been found to inhibit increase in body weight and visceral fat content in high-fat diet-fed mice (Hamao *et al*, 2011). Particularly, the n-butanol (BuOH)-soluble fraction of the extract and its primary component, chakasaponins II, was found to reduce food intake in high-fat as well as normal diet-fed mouse. They have also been found to inhibit the levels of neuropeptide Y (NPY) mRNA in the hypothalamus. Moreover, chakasaponins II were found to release 5-hydroxytryptamine (5-HT) from the isolated ilea cells of mice, thus suppressing appetite signals.

Hypoglycemic

Hypoglycemic properties of tea has been well known for ages. Lately, the anti-diabetic properties of tea flowers have also been extensively worked upon and eventually revealed. Matsuda *et al* (2012) demonstrated the role of chakasaponins I–III (50 and 100 mg/kg) extracted from tea flower buds in significantly lowering glucose levels in olive oil or sucrose-loaded mice. Blood glucose levels in alloxan-induced diabetic mice was found to decline considerably by continuous oral administration of tea flower polysaccharides (Han *et al*, 2011a). Similar hypoglycemic effect of tea flower polysaccharides were observed in diabetic Sprague-Dawley mice induced by alloxan (Cai *et al*, 2011). Methanolic and n-butanol soluble fractions of tea flowers have been found to possess suppressive effects on serum glucose levels in sucrose-loaded mice. Specifically, floratheasaponins A, B and C present in the n-

butanol soluble fraction have been shown to have hypoglycemic properties (Yoshikawa *et al*, 2008).

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