

# Histidine: A Systematic Review on Metabolism and Biological Affecting on Human body

Aseel H. Abad Al-Ameer

Department of Chemistry, College of Science, University of Baghdad, Baghdad, Iraq

## Abstract:

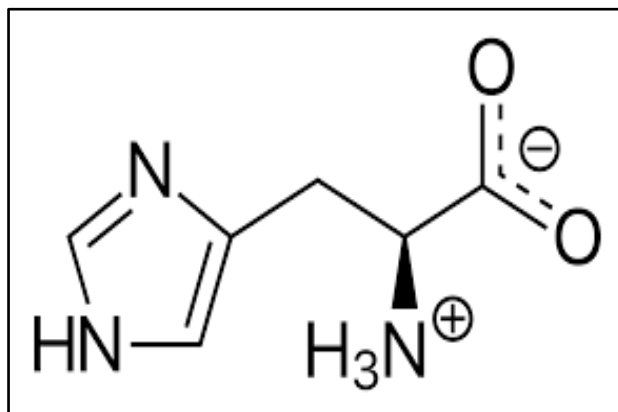
Histidine is a fundamental amino acids (EAA) in warm blooded animals, poultry and fish. We wish to give an outline of the digestion and physiological properties of histidine in people through a standard survey keeping the PRISMA's Instructions (Preferred Reporting Items for Systematic Reviews and Meta-Analyses). In people, His. From the diet might be connected With functions that do create metabolic condition and affects particle ingestion. In rodents, histidine supplementation rises food consumption. It in addition offers neuroprotection in an installation underlying time and could keep versus Epileptic convulsions. Histidine is primarily important in chickens as a controlling component for the amalgamation of carnosine, and has great enemies of oxidant action. Dietary histidine in fish is known to be the single most essential factor to stay away from waterfalls. His is a special item for the development of milk protein through ruminants and may be the main limiting AA for advancement. In addition, levels. Hist. supplementation may be to blame for people's feeding and memory inconvenience and can persuade development hindrance and metabolic disorder in the right form. To decide, the cravings Hist, as with the other EAA, was used for coming about because of development and AA compliance in tissues and furthermore have definite metabolic jobs expect species and nutritionry

**Keywords:** metabolic rate, physiological properties, humans, animal classes

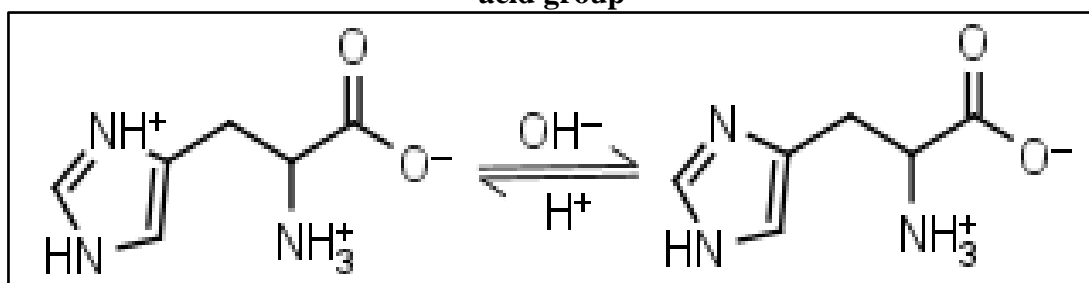
## Introduction:

Histidine (His) [1] is  $\alpha$ -amino corrosive used in the biosynthetic pathways of proteins. It was first disengaged from salmon protamine by Albrecht Kossel in 1896 [2] He picked the name histidine from the Greek word histion signifying "tissue." It is a fundamental amino corrosive with an imidazole side chain It comprise of a  $\alpha$ -amino gathering (which is in the protonation of  $-\text{NH}_3^+$ , a carboxylic corrosive crowd (In the deprotonated COO structure under organic conditions) and an imidazole sideways chain (mostly framed protonated), inventoring it at physiological pH as an emphatically charged amino corrosive. During important only for infants, right off the bat during important It has now been added to be fundamental for developed individuals in longer-term readings as well.[3] It is regulated by the CAU and CAC codons. Histidine The first was protected in 1896 by German doctor Albrecht Kossel and Sven Gustaf Hedin. [4] It is also a pioneer in histamine, an important fiery expert in resistant reactions. The imidazole side chain's corrosive base assets was associated with the synergistic mechanism of various enzymes.[5] The essential N2 of the histidine synopsis from serine, threonine, or cysteine proton to serine invigorates it as a nucleophile in reactant groups of three. It is used in a histidine proton vessel to transfer protons rapidly. In the abstraction of the proton, it can do this essential N2 to type an emphatically charged temporary and afterward practice another particle, a cushion, To generate the proton out of its acidic nitrogen, imidazole.. Histidine sorts edifices with various metal particles. side chain of the histidine leftover portion regularly fills in In metal proteins Just as a ligand. In myoglobin and hemoglobin, one paradigm is the hub base based on Fe. For protein purification, poly-histidine names (of at least six back to back H buildups) are used with micromolar partiality by authoritative sections of Ni or Co. [6] Standard poly-histidine peptides present in snake toxin *Atheris squamigera* have been shown to bind

Zn(2+), Ni(2+) and Cu(2+) and touch toxin metalloprotease ability. [7] In comparison, low histidine-rich-multifaceted nature districts metal is contained in the -official and particularly nickel-cobalt restricting proteins



**Figure 1. Histidine structure: histidine (HIS) contains an  $\alpha$ -amino group, a carboxylic acid group**



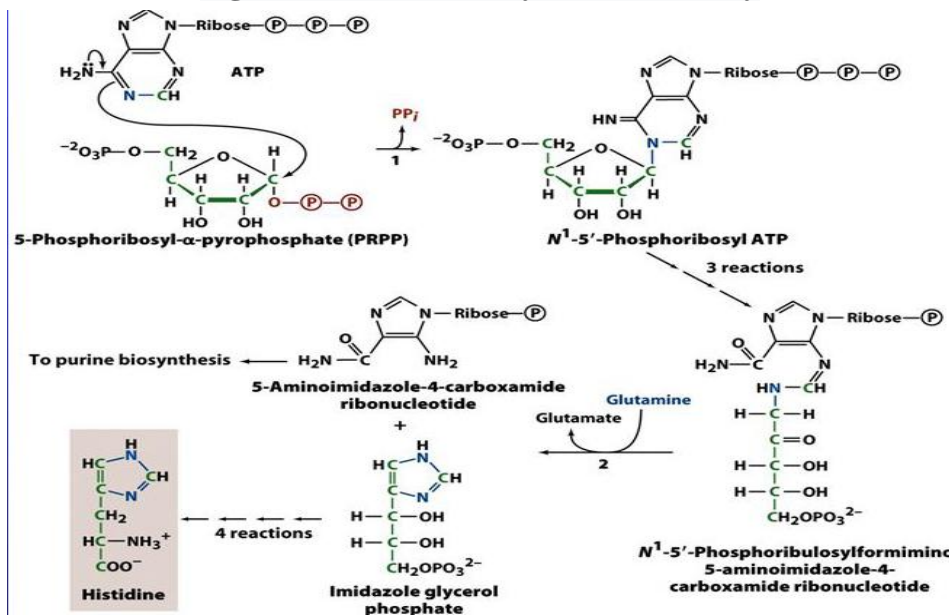
**Figure 2. Acid-base properties of the imidazole side chain**

## 1- Collective Physiological Characters of Histidine in Various Classes

### 1-1 Biosynthesis

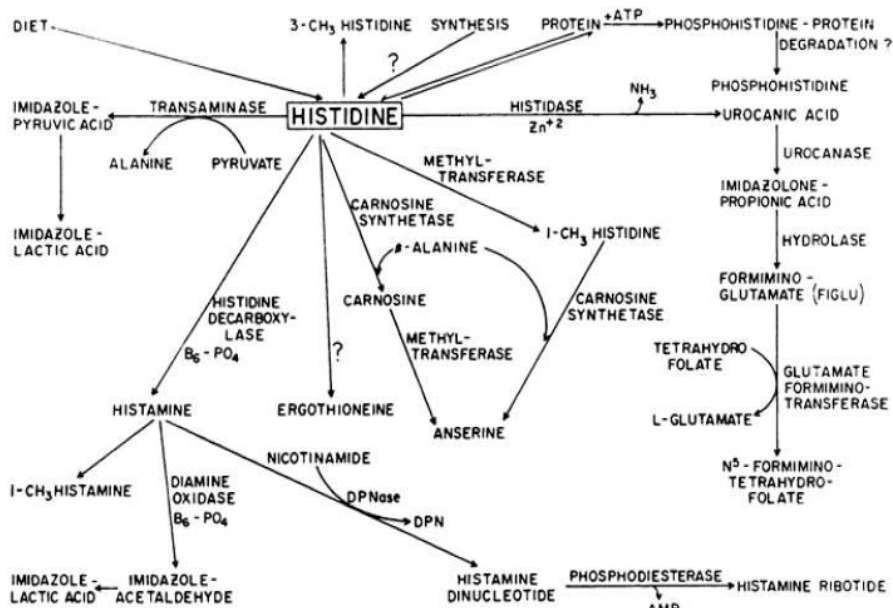
Histidine is produced from phosphoribosyl pyrophosphate (PRPP), which is pentose-phosphate formed by ribose-5-phosphate by ribose-phosphate diphosphokinase from ribose-5-phosphate. The basic response of its biosynthesis This is the aggregation of PRPP and adenosine triphosphate by the catalyst ATP-phosphoribosyl transferase (ATP).His appears with ATP-phosphoribosyl transferase in the image.[8] The phosphoribosyl-ATP, phosphoribosyl-AMP (PRAMP) build-up element, which is an irreversible step, is hydrolyzed at that stage by the His4 material. At that point, His4 catalyzes the development of phosphoribosylformiminoAICAR-phosphate, which is then changed to phosphoribulosylformimino-AICAR-P by producing the His6 material. [9] His7 parts phosphoribulosylformimino-AICAR-P to The D-erythro-imidazole-glycerol-phosphate structure. His3 then structures the water stream of imidazole acetol-phosphate. His5 labels L-histidinol-phosphate at that point, which is then hydrolyzed by His22. building histidinol. His4 catalyzes the oxidation of L-histidinol to frame L-histidinal, an amino aldehyde. In the keep going advance, L-histidinal is changed to L-histidine.[10][11]

**Figure 3. Histidine Biosynthesis Pathway**



### 1-2. Metabolism

Histidine endures changed metabolic ways (Figure 4). It may quite well be methylated to either 1-methyl or 3-methyl histidine, or changed to imidazole-pyruvic corrosive by transaminase, which creates corrosive imidazole-lactic by imidazole-lactic corrosion.decreasing. Fetal rodent livers and misfortunes after birth are packed with transaminase. Its action can be changed by food: when diet protein is reached, it increases above 25%, though it isn't changed underneath 25%. Histidine can be abbreviated to frame carnosine and anserine with  $\beta$ -alanine, which is incorporated in defense similar to oxidative strain. [12]. In order to deliver histamine, it may either be thick with decarboxylation or it may undergo permanent rot. Histidine alkali lyase, Also referred to as histidase, which is a cytoplasmic chemical[13], the alpha-amino collection of L-histidine is far away[14] to do this. It is a non-oxidative, permanent deamination targeted at the production of corrosive and odor salts that are trans-urocanic. This is the major process of histidine corruption in well-developed mammals and bacteria.



**Figure 4. HistidineMetabolism pathway**

### 1-3.Histidine Catabolism

Ordinary every day limit of In established persons, HIS of about 800 mg indicates that the comparable volume of HIS should be pillaged. The core route of HIS catabolism (Figure 5) begins with histidase-catalyzed deamination (EC 4.3.1.3), leading to the production of transurocanate and odorous salts. The compound is located primarily in the corneum layer of the skin and liver.

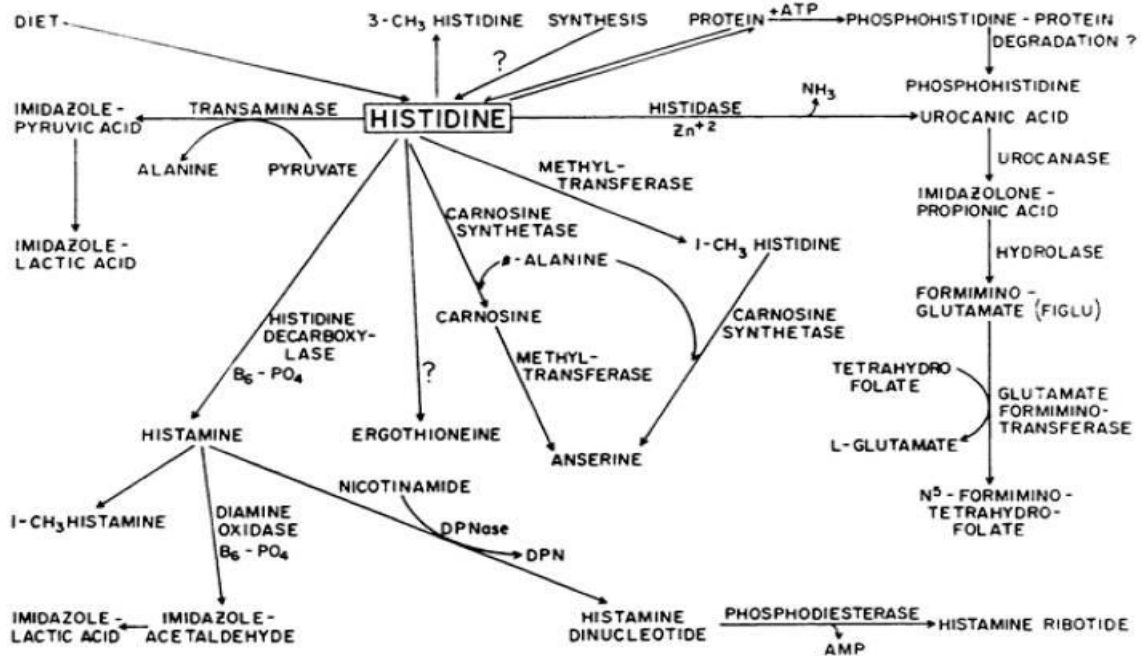


Figure 5. Histidine catabolism pathway

#### 1.3.1.Histidine Catabolism in the Skin

Filaggrin, a skin section protein that contains amazing HIS, is the primary source of histidase in the skin to create alkali and urocanate. [15]. Explanations behind this the greater part of the alkali shaped In the splanchnic area If urea in the liver is detoxified, the skin in the fundamental flow should be regarded as an essential premise for blood-smelling salts. Since urocanase (the second protein in HIS catabolism) is needed by the skin, transurocanate amasses in the corneum layer, contributing to the creation of "normal saturating components" and activities as perhaps the main bright (UV) operation. retaining mixes [16]. Transurocanate is isomerized into cis-urocanate in the presence of UV radiation, which may impair the UV radiation-induced concealment of the resistant structure [17].

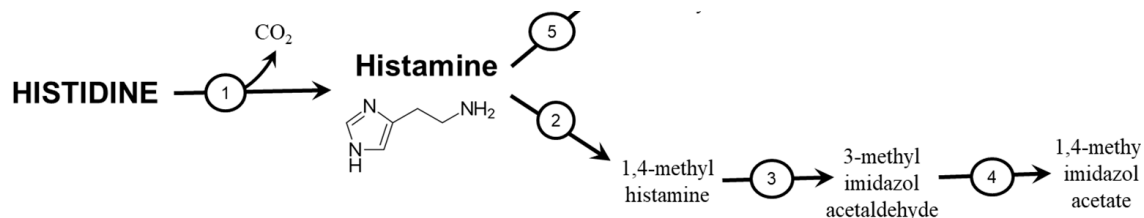
#### 1.3.2.Histidine Catabolism in the Liver

In the liver, articulation of histidase is constrained by the availability of HIS. Movement of histidase improves when protein admission is in stature and reduces when protein admission is low. [The 18,19]. Urocanase (EC 4.2.1.49) changes the urocanate formed by an imidazolone propionic corrosive histidase reaction in the liver that is hydrolysed to formiminoglutamate (FIGLU). In tetrahydrofolate (THF) by formimino transferase (E.C. 2.1.2.5)-subordinate reaction, FIGLU is converted to glutamic corrosive. FIGLU collects and HIS catabolism is hampered on the off risk that folate is insufficient. [20]; HIS-stacking (FIGLU discharge test) is an indicative instrument for THF nonattendance [21]. Decrease of THF can trigger an unmistakable decrease in the mass of the serine glycine blend after HIS filling (see Figure 3), as defined by Meléndez-Hevia et al.[22] and Hole'cek and Vodeni'carovová [23]. From some sources, THF is subordinate that may influence the transformation of HIS through the HIS hardship process (Figure 3). Several articles have

established that by growing the achievable content of THF[24,25], methionine, The complex catabolism of HIS is induced by methionine, S-adenosylmethionine, homocysteine, and S-adenosylhomocysteine.S-adenosylmethionine, homocysteine, and S-adenosylhomocysteine generate complex HIS catabolism. Glutamate formed by the formimino transferase reaction may be intended for a glutamine mixture, may shift to deaminated-ketoglutarate in a glutamate dehydrogenase reaction, or may be delivered to the blood. [26]. Due to additional alanine fixation achieved from extrahepatic tissues, transamination to alanine is unlikely to occur. A strong concern of HIS extends HIS transformation into the pathway of HIS corruption, resultant an expanded alkali fabricate and changed centralizations of some amino acids, generally expanded convergences of glutamate, alanine, and glutamine and diminished stretched chain amino acids (BCAA) focuses in the blood plasma [27].

#### 1.4. Histamine

In pole cells and basophils, from which it is extracted, the most prominent histamine is mixed and deposited in granules delivered by means of degranulation brought by immunological incitement (Figure 6), chiefly associations Allergens with antibodies to IgE. In the mind, The extra vital spots of histamine combining and pressing are parietal cells in the stomach and histaminergic neurons. Hydrochloric corrosive products of parietal cells; histaminergic back nerve core neurons regulate a variety of physiological occupations, containing craving, alertness, feelings, and intellectual capacities. Histamine is planned L-histidine decarboxylase (EC 4.1.1.22) starts in different tissues with HIS decarboxylation.. 1,4-methyl imidazoleacetic corrosive is corrupted into unhindered histamine; oxidation of imidazoleacetic corrosive is the other direction to histamine debasement. The transfer of metabolites is to the urine or other metabolites are handled by: 1, histidine decarboxylase; 2, Histamine-N-methyltransferase; 3, monoamine oxidase; 4, aldehyde dehydrogenase; 5, diamine oxidase.



**Figure 6. Synthesis and degradation of histamine**

Histamine shows likewise a fundamental character As a regulator of muscle microcirculation by exercise and continuous vasodilation after rehearsal, [28,29]. The appearance of histidine decarboxylase is energized by cytokines (especially IL-1 and TNF-) in polar cells, vascular endothelial cells, and muscle strands themselves. more noteworthy than before Temperature, reduced pH and inducible factor 1 for hypoxia [30-33]. In invulnerable tissues, the brain, and the cerebrum, dietary HIS raises histamine concentrations[34-37] on histamine statures granted to some exercises.Modified capacity of the resistant course of action, hypersensitive responses, or potentially peptic ulcers have not been depicted after HIS organization. All things considered, HIS association has been appearing to influence cerebrum work [38-46].

#### 2. Effect of Histidine Level and Supplementation in Humans Bodies

In line with the FAO, the day-to-day commitment to histidine in developing individuals is 8 to 12 mg/kg of body weight per day. The usual consumption of histidine was observed to be in the range of 2.12 and 2.40 g daily (i.e. around 30 to 35 mg/kg body weight daily) in

normal, grown-up feeding regimens in Europe, the United States and Japan, where the 99th percentile intake was 5.20 g/day in men aged 50-70 years[47, 48].

### **2.1. Histidine Intake and Consequences on Body Histidine Status**

Steele and Le Bovit[49] arranged histidine tolerance in females who expected two degrees of L-histidine, 1.5 and 5 g/day. Evaluations showed that the 1.5 g section increased the plasma histidine 2-to 8-overlap at thirty minutes after consumption, while the 5 g section boosted the 7-to-11-crease ratio above the fasting mark. The fulfilled plasma histidine remained elevated for a brief amount of time in situations (1.5 and 5 g/day) and gradually decreased before the fasting level was reached. decrease of the red platelet histidine was a lot requiring additional time than that of plasma. Square and associates[50] have arranged histidine resilience in three adults in fine fettle (two females and one male) who produced 5 g of free histidine in 100 ml of softened water. Blood testers were conducted at one, two and four-hour intervals. The amount of histidine in the plasma increased an hour after assimilation of 5 g of histidine to around 7.5, 12.5 and several times the fasting level. On both examinations, the plasma histidine amount was not precisely for the one-hour test at two-hour intervals and persistent to declining. Be that as it may. By the end of the resilience test, it did not return to the fasting point. These two readings have demonstrated that extraordinary levels of histidine quickly inducing a significant increase in histidine plasma, which then falls in a couple of hours.

### **2.2. Histidine Supplementation and Eating Disorders**

Henkin[51] found that points of anorexia unexpectedly began to battle within four to six days after differing amounts of histidine supplementation in healthy, youthful school men. For volunteers receiving 8 g of histidine a day, anorexia and misfortunes were high, weighing between 2-3 lbs. over a time of multi week, regardless of consistent reassurance to complete their dinners. All things considered, the best-suffered feast was breakfast, followed by lunch, and dinner was the least-suffered supper. Continuing to be correlated with higher histidine measurements, 16 and 32 g was consistently associated with histidine. Both participants developed a decline in taste sharpness (hypogeusia) and eventually a decrease in the sense of smell (hyposmia). The enhancement of taste mutilation (dysgeusia) and smell comprehension was correlated with further association of higher segments of histidine (dysomnia). Okubo and Sasaki [52] broke down the association between's Between 1689 Japanese female understudies, dietary histidine and energy consumption using an objective routine background survey of feeding. Results revealed that the extent of histidine to protein (histidine/protein) was released from other dietary products and was contradictory and ultimately related to energy confirmation. Likewise, Nakajima and accomplices[53] perceived a negative association between histidine/protein and energy intake in 26 male and 38 female understudies ( $r=-0.18$  in males and  $r=-0.34$  in females,  $p<0.05$ ). [54] dealt with eight sound men (ages 32 to 38) with 4 g of histidine supplement each day for about a month. Results revealed that this entirety has little profound effects on wisdom of wanting, taste or scent, diet affirmation, or body weight. the Norwegian Logical Board of trustees for Sanitation (Holvik et al., 2016) believed that 4 to 4.5 g/day supplementation does not have antagonistic effects in humans, identifying 57 mg/kg of body weight a day for a 70 kg adult.

### **2.3. Histidine and Memory Disorders**

Geliebter et al. examined the influence of stable measurements of his 24 to 64 g, combined with crushed orange, for approximately one month on strong subjects [55]. Two participants Anguishing sensations in their eyes and some concentration discomfort were recorded by those who received 64 g a day. [57] showed that the use of histidine (1.65 g/day) actually declined step by step vibes of exhaustion, extended adequacy. When executing recall

functions, in subjects with elevated weariness and rest interference scores, unflinching wisdom and concentration advanced.

#### **2.4. Histidine and Metal Ion Status**

The effect of histidine on molecule maintenance was investigated in a couple of assessments. For eg, iron maintenance in 113 subjects was not impaired by supplementation with histidine (416 to 2080 mg/day) [58]. Zinc histidine structures, retained over zinc sulfate in ten sound subjects, were preferred in another evaluation. Subsequently, the consumption of 1:2 or 1:12 zinc frameworks with histidine extended serum-zinc obsession by 25 percent more than the intake of zinc sulfate. No improvements in the release of zinc were noticed in this evaluation. [59]. In 1980, Henkin what's more, partners considered the effects of assessed augmentations of histidine partitions (8.1–64.8 g, step by step). With healthy volunteers as in people with scleroderma. They're going to saw an assessed rise in the release of urinary zinc. Urinary zinc release extended from  $454 \pm 50 \mu\text{g}$  consistently to  $5269 \pm 840 \mu\text{g}$  consistently behaved uniquely in comparison to the control condition in subjects obtaining high sections of histidine. Generally, in particular subjects with. The greatest proportion of histidine supplementation (64.8 g step-by-step), 0.5% of the measured. The entire body's zinc reservoir was depleted by peeing every day. A neurodegenerative disease is the Menkes ailment (MD), that can be achieved by improvements in the consistency of ATP7A and can incite destruction in youth. Indications. The loss of production of Cu-subordinate proteins is due to this pollution (cytochrome oxidase, tyrosinase, dopamine beta hydroxylase, superoxide dismutase, lysyl oxidase, and sulfhydryl oxidase). Copper therapy in the presence of MD has been shown to be unsuccessful in patients. Inquisitively, copper histidine treatment is important in pivoting skin and hair improvements, enhancing joint tone, socio-scholarly achievements, weight gain, and insusceptibility due to the chelating sway [60-62].

#### **3. Effect of Histidine on Mineral Metabolism**

Histidine, as seen at a late stage, can chelate metal particles. The effect and assistance of histidine confirmation on iron, copper and zinc absorption has been investigated. in a few evaluations. Histidine supplementation has not impacted tissue iron levels and fecal iron mishaps [63]. It expanded osmosis right where histidine was applied to a  $^{59}\text{Fe}$  schedule containing ascorbic destructive osmosis [64]. This indicates some rapid Reaction of iron and histidine and is consistent with the principle of identifying and then swallowing an AA-iron chelate. No influence of histidine affirmation has been found in rodents with deference to [65]. Histidine was used to energize the take-up of copper in the hepatic, placental, and neuronal contacts in vitro [66-69]. The impact of histidine-profiting on zinc status has been studied by a couple of assessments. The impact of supplementation of histidine is debatable. Taylor and Freeman. [70] Investigated the effects of both severe (250 mg/hr histidine intravenous imbuement) and consistent (500 mg/day 43-day gavage administration) histidine supplementation. After intense and consistent histidine interaction, they discovered an extension of zinc release by three to multiple occasions. Elsewhere, the loss of plasma zinc was seen in extraordinary, but not tenacious, supplementation in any case. In energetic adult rodents, dietary repletion with zinc chloride fortified with L-histidine (40 mg/kg of diet) was all the more notable when scholarly deficiency was exchanged due to zinc loss rather than zinc salt repletion alone. The Action of the  $^{65}\text{Zn}$  vehicle over the endothelium of the cerebrum when histidine was applied to perfusion may explain this impact. [71]. Snedeker and Greger [72] observed that the preservation and use of zinc in the feeding regimen was much more severely impaired by protein levels than by histidine levels. In any event, various tests are nitty gritty that histidine can cause a lack of zinc. In rodents dealt with histidine levels in a way that is better than 4 g/kg BW/A fundamental decline in plasma zinc was

observed on the day from seven to 46 weeks. Specifically, rat diet supplementation with 50 g/kg of histidine did not alter zinc turnover velocity ( $^{65}\text{Zn}$  turnover velocity from 2 to roughly 1 month after lone tracer imbue), but the feeding regimen increased with 8% of histidine, triggering an outrageous need for zinc (half the plasma zinc content decreased) [73]. This effect of supplementation with histidine on the status of zinc may depend on dietary zinc affirmation. Histidine supplementation, of course, did not encourage any movements in the zinc state as rodents were fed with zinc-sufficient feeding regimen (zinc obsessions,  $^{65}\text{Zn}$  tissue course, and tissue-unequivocal activities). In either event, with low zinc evidence, histidine supplementation resulted in Help associated with extended fecal release and a more limited normal half-life [74] is lower than  $^{65}\text{Zn}$ .

#### **4. Histidine and Cancer**

Late results indicate that catabolism and confirmation of histidine affects the methotrexate affectability of disease cells [75]. For some solid tumors and blood-threatening developments, methotrexate is an anticancer therapy, but can be dangerous for separate non-sickness cells. The usage of Cyclodeaminase formimidoyltransferase (FTCD), histidine salt lyase (HAL), and area 1-containing amidohydrolase (AMDHD1). CRISPR/Cas9 lowered the cell affectability of methotrexate. Methotrexate and histidine therapy in mice, by contrast, caused a controlled decrease in tumor size, which was considerably more significant than in all other treatment assistants. Taken together, these findings demonstrate the role of the histidine corruption pathway in the ability of chemotherapy specialist methotrexate [76].

#### **5. Toxicity Dose**

In mice, destructiveness divisions were transferred from substitute perspectives: their carcinogenicity was close to unsafe measurements. During the 13 week duration during which different quantities of histidine were administered to rats (containing 0, 3.1, 6.2, 12.5, 25 and 50 g/kg of histidine), no death or terrible consequences were reported. [77]. another evaluation studied the hurtfulness and malignancy associated with the Histidine characteristics in 100 rats for 104 weeks [78]. Two social cases, 50 people and 50 women, concerned with calorie counts containing 0, 12.5 and 25 g/kg histidine, were included in this evaluation. They saw that tumors were created in any case. one of the three social affairs. Regardless, these tumors were identical to those that fill steeply in F344 rodents. No basic differentiations between these three social occasions, a recurrence or form of tumor was seen. The manufacturers considered that, under their exploratory conditions, histidine was not causing disease in these rodents. Accomplices and Gullino [79] mulled over harmfulness of AAs. They pondered the DL50, Histidine, the section that kills half the rodents, and the part that eliminates all the mice is DL99.9. The findings of Noxiousness revealed that the DL50 body weight was 23 mmol/kg and the DL100 body weight was 33 mmol/kg. The bit was not differentiated between the two isomers, L-histidine and D-histidine.

#### **6. Risks and Side Effects**

While it's incomprehensible that you'd consume high proportions of histidine from sustenances alone, it's possible to consume excess aggregates from supplements, which can cause certain outcomes. Studies have found that when people take amazingly high measurements of histidine, around 32 grams/day or more, they can experience results like muscle deficiency, sluggishness and shortcoming, cerebral torments, stomach related issues like squeamishness and loss of yearning, agony, and powerless memory. A part of these may be a direct result of negative nitrogen balance. Other antagonistic effects appended to high histidine levels have similarly been showed up in animal ponders, yet it's dark how these effects reach out to individuals. In examinations including rodents, troubles joined to high



histidine levels in the brain and liver have included copper deficiency, reduced liver limit, raised cholesterol and loss of wanting. A segment of the possible consequences of eating up an abundance of protein generally speaking join weight procure, kidney issues, deterrent and horrendous breath. Anyone with kidney or liver contamination should not consume a ton of amino acids without working with an expert [80-82]

## Conclusions

His. is a key AA and its essentials have been obtained from tissue enhancement and AA synthesis. Despite its work in protein assimilation as a beneficial AA, histidine has unique metabolic roles. As a result, histidine is limited to consuming while routine tantamount to the necessity for advancement, It is possible to link its supplementation with an unparalleled show. Right when this supplementation gives histidine past advancement essential, various focal points This paper can be considered exhaustively as depicted. In either case, when histidine is given in abundance by this supplementation, AA ponderousness will cause the production of a decrease and food affirmation and other undesirable consequences. Beginning there, these section results of histidine are summed up:

1. The step-by-step need for histidine in humans is 8 to 12 mg/kg of body weight per day in adults and 8 to 12 mg/kg of body weight per day in adults typical confirmation in like manner grown-up eating regimens About 30 to 35 mg/kg body weight per day was seen in Europe, the USA and Japan (2.12 and 2.40 g consistently). Move by step-by-step supplementation of histidine, below 2 g/consistently, increases disruption in overweight and husky individuals, decreases vibes of exhaustion, and fabricates obsession and capability at work. A step by step In atopic dermatitis infection, the affirmation of 4 g reduces fact. Extraordinary anorexia is instigated if the supplementation rises above 8 g of histidine. Finally, from 16 to 64 g/day, supplementation in a large proportion produces a reduction in the sharpness of taste and fragrance and the onset of headaches, insufficiency, sleepiness, squeamishness and trouble with memory.

2. In rodents, up to 25 g/kg of dietary supplementation with histidine is essential. A significant portion of 50 g/kg of diet may have unmistakable threatening consequences, such as a colossal drop in healthy Body weight, food intake, and a growth buffer. As a result of this adverse effect, the cap of the respectable portion of histidine was set at 25 g/kg of routine consumption.

Histidine supplementation, depending on the species, can be charming with multiple backgrounds to wrap up. In humans, data indicate Supplementation of histidine as a device for increasing metabolic characteristics, skin dysfunctions, and memory of both therapeutic and stimulating interests. Histidine supplementation in animal processing will bear some enormity with development of both the yield and quality of potential effects. Histidine's criticality and conglomerating In various physiological limits, proof of its centrality makes histidine a phenomenal AA that merits additional interest in human and animal sustenance.

## References:

1. ME. Albrecht Kossel, a biographical sketch. Yale J Biol Med. 1953;26:80-97.
2. <http://prowl.rockefeller.edu/aainfo/solub.htm> <sup>[full citation needed]</sup>
3. "Nomenclature and Symbolism for Amino Acids and Peptides". IUPAC-IUB Joint Commission on Biochemical Nomenclature. 1983. Archived from the original on 9 October 2008. Retrieved 5 March 2018.
4. Kopple, J D; Swendseid, M E (1975). "Evidence that histidine is an essential amino acid in normal and chronically uremic man". Journal of Clinical Investigation. **55** (5): 881-91. doi:10.1172/JCI108016. PMC 301830. PMID 1123426.

5. Vickery, Hubert Bradford; Leavenworth, Charles S. (1928-08-01). "ON THE SEPARATION OF HISTIDINE AND ARGININE IV. THE PREPARATION OF HISTIDINE" (PDF). *Journal of Biological Chemistry*. **78** (3): 627–635. ISSN 0021-9258
6. ngle, Robert A. (2011). "Histidine Biosynthesis". The Arabidopsis Book. **9**: e0141. doi:10.1199/tab.0141. PMC 3266711. PMID 22303266.
7. Roberts, John D. (2000). *ABCs of FT-NMR*. Sausalito, CA: University Science Books. pp. 258–9. ISBN 978-1-891389-18-4.
8. Bornhorst, J. A.; Falke, J. J. (2000). "Purification of proteins using polyhistidine affinity tags". *Methods in Enzymology*. **326**: 245–254. doi:10.1016/s0076-6879(00)26058-8. ISSN 0076-6879. PMC 2909483. PMID 11036646.
9. Watly, Joanna; Simonovsky, Eyal; Barbosa, Nuno; Spodzieja, Marta; Wieczorek, Robert; Rodziewicz-Motowidlo, Sylwia; Miller, Yifat; Kozłowski, Henryk (2015-08-17). "African Viper Poly-His Tag Peptide Fragment Efficiently Binds Metal Ions and Is Folded into an  $\alpha$ -Helical Structure". *Inorganic Chemistry*. **54** (16): 7692–7702. doi:10.1021/acs.inorgchem.5b01029. ISSN 1520-510X. PMID 26214303.
10. Ntountoumi, Chrysa; Vlastaridis, Panayotis; Mossialos, Dimitris; Stathopoulos, Constantinos; Iliopoulos, Ioannis; Promponas, Vasilios; Oliver, Stephen G; Amoutzias, Grigoris D (2019-11-04). "Low complexity regions in the proteins of prokaryotes perform important functional roles and are highly conserved". *Nucleic Acids Research*. **47** (19): 9998–10009. doi:10.1093/nar/gkz730. ISSN 0305-1048. PMC 6821194. PMID 31504783.
11. Alifano, P; Fani, R; Liò, P; Lazcano, A; Bazzicalupo, M; Carlomagno, M S; Bruni, C B (1996-03-01). "Histidine biosynthetic pathway and genes: structure, regulation, and evolution". *Microbiological Reviews*. **60** (1): 44–69. doi:10.1128/MMBR.60.1.44-69.1996. ISSN 0146-0749. PMC 239417. PMID 8852895.
12. Jump up to:<sup>a b</sup> Kulis-Horn, Robert K; Persicke, Marcus; Kalinowski, Jörn (2014-01-01). "Histidine biosynthesis, its regulation and biotechnological application in *Corynebacterium glutamicum*". *Microbial Biotechnology*. **7** (1): 5–25. doi:10.1111/1751-7915.12055. ISSN 1751-7915. PMC 3896937. PMID 23617600.
13. Adams, E. (1955-11-01). "L-Histidinal, a biosynthetic precursor of histidine". *The Journal of Biological Chemistry*. **217** (1): 325–344. ISSN 0021-9258. PMID 13271397
14. Taylor R.G., Levy H.L., McInnes R.R. Histidase and histidinemia. *Clinical and molecular considerations*. *Mol. Biol. Med.* 1991;8:101–116. [PubMed] [Google Scholar]
15. Mehler A.H., Tabor H. Deamination of histidine to form urocanic acid in liver. *J. Biol. Chem.* 1953;201:775–784. [PubMed] [Google Scholar]
16. Tessari P. Nonessential amino acid usage for protein replenishment in humans: a method of estimation. *Am J Clin Nutr* 2019;110:255–64.
17. Coltorti M, Di Simone A, Budillon G. Histidase and urocanase activities of liver and plasma. Correlations between tissue enzyme levels and plasmatic increases during human and mouse viral hepatitis. *Clin Chim Acta* 1966;13:568–73.
18. Steinert, P.M.; Cantieri, J.S.; Teller, D.C.; Lonsdale-Eccles, J.D.; Dale, B.A. Characterization of a class of cationic proteins that specifically interact with intermediate filaments. *Proc. Natl. Acad. Sci. USA* **1981**, *78*, 4097–4101. [CrossRef]
19. Hug, D.H.; Hunter, J.K.; Dunkerson, D.D. The potential role for urocanic acid in sunlight in the immune suppression associated with protein malnutrition. *J. Photochem. Photobiol. B.* 1998, *44*, 117–123. [CrossRef]
20. Kang-Lee, Y.A.; Harper, A.E. Effect of induction of histidase on histidine metabolism in vivo. *J. Nutr.* **1979**, *109*, 291–299. [CrossRef] [PubMed]

21. Torres, N.; Martínez, L.; Alemán, G.; Bourges, H.; Tovar, A.R. Histidase expression is regulated by dietary protein at the pretranslational level in rat liver. *J. Nutr.* **1998**, *128*, 818–824. [CrossRef] [PubMed]
22. Luhby, A.L.; Cooperman, J.M.; Teller, D.N. Urinary excretion of formiminoglutamic acid: Application in diagnosis of clinical folic acid deficiency. *Am. J. Clin. Nutr.* **1959**, *7*, 397–406. [CrossRef] [PubMed]
23. Silverman, M.; Gardine, R.C.; Condit, P.T. A method for the detection of N-formiminoglutamic acid in urine. *J. Natl. Cancer Inst.* **1958**, *20*, 71–77.
24. Meléndez-Hevia, E.; De Paz-Lugo, P.; Cornish-Bowden, A.; Cárdenas, M.L. A weak link in metabolism: The metabolic capacity for glycine biosynthesis does not satisfy the need for collagen synthesis. *J. Biosci.* **2009**, *34*, 853–872.
25. Fell, D.; Steele, R.D. Effect of methionine on in vivo histidine metabolism in rats. *J. Nutr.* **1983**, *113*, 860–866. [CrossRef]
26. Billings, R.E.; Noker, P.E.; Tephly, T.R. The role of methionine in regulating folate-dependent reactions in isolated rat hepatocytes. *Arch. Biochem. Biophys.* **1981**, *208*, 108–120. [CrossRef]
27. Cynober, L. Metabolism of dietary glutamate in adults. *Ann. Nutr. Metab.* **2018**, *73*, 5–14. [CrossRef]
28. Holeček, M.; Vodeničarovová, M. Effects of histidine load on ammonia, amino acid, and adenine nucleotide concentrations in rats. *Amino Acids* **2019**, *51*, 1667–1680.
29. Greaves MW, Sabroe RA. Histamine: the quintessential mediator. *J Dermatol* 1996;23:735–40.
30. Andersson K, Chen D, Mattsson H, Sundler F, Hökanson R. Physiological significance of ECL-cell histamine. *Yale J Biol Med* 1998;71:183–93.
31. Haas HL, Sergeeva OA, Selbach O. Histamine in the central nervous system. *Physiol Rev* 2008;88:1183–241.
32. Watanabe T, Taguchi Y, Shiosaka S, Tanaka J, Kubota H, Terano Y, Tohyama M, Wada H. Distribution of the histaminergic neuron system in the central nervous system of rats; a fluorescent immunohistochemical analysis with histidine decarboxylase as marker. *Brain Res* 1984;295:13–25.
33. Prell GD, Hough LB, Khandelwal J, Green JP. Lack of a precursor-product relationship between histamine and its metabolites in brain after histidine loading. *J Neurochem* 1996;67:1938–44.
34. Yoshikawa T, Nakamura T, Yanai K. Histamine N-methyltransferase in the brain. *Int J Mol Sci* 2019;20:737.
35. Lee, N.S.; Fitzpatrick, D.; Meier, E.; Fisher, H. Influence of dietary histidine on tissue histamine concentration, histidine decarboxylase and histamine methyltransferase activity in the rat. *Agents Actions* **1981**, *11*, 307–311. [CrossRef]
36. Martin, S.K.; Harmon, D.L.; Conway, C.E.; Vanzant, E.S.; McLeod, K.R. Influence of dietary histidine on basophil release, circulating concentration, and urinary excretion of histamine in domestic felines. *J. Appl. Res. Vet. Med.* **2012**, *10*, 289–299.
37. Lozeva, V.; Tarhanen, J.; Attila, M.; Männistö, P.T.; Tuomisto, L. Brain histamine and histamine H3 receptors following repeated L-histidine administration in rats. *Life Sci.* **2003**, *73*, 1491–1503. [CrossRef]
38. Yoshikawa, T.; Nakamura, T.; Shibakusa, T.; Sugita, M.; Naganuma, F.; Iida, T.; Miura, Y.; Mohsen, A.; Harada, R.; Yanai, K. Insufficient intake of L-histidine reduces brain histamine and causes anxiety-like behaviors in male mice. *J. Nutr.* **2014**, *144*, 1637–1641. [CrossRef] [PubMed]
39. Kasaoka, S.; Tsuboyama-Kasaoka, N.; Kawahara, Y.; Inoue, S.; Tsuji, M.; Ezaki, O.; Kato, H.; Tsuchiya, T.; Okuda, H.; Nakajima, S. Histidine supplementation suppresses

- food intake and fat accumulation in rats. *Nutrition* **2004**, 20, 991–996. [[CrossRef](#)] [[PubMed](#)]
40. . Goto, K.; Kasaoka, S.; Takizawa, M.; Ogawa, M.; Tsuchiya, T.; Nakajima, S. Bitter taste and blood glucose are not involved in the suppressive effect of dietary histidine on food intake. *Neurosci. Lett.* **2007**, 420, 106–109. [[CrossRef](#)] [[PubMed](#)]
  41. . Agwara, M.O.; Ndifon, P. T.; Ndosiri, N. B.; Paboudam, A. G.; Yufanyi, D.M.; Mohamadou, A., *Bulletin of the Chemical Society of Ethiopia*", (2010), 24(3), 383–389.
  42. . Asahi, R.; Tanaka, K.; Fujimi, T.J.; Kanzawa, N.; Nakajima, S. Proline decreases the suppressive effect of histidine on food intake and fat accumulation. *J. Nutr. Sci. Vitaminol. (Tokyo)* **2016**, 62, 277–280. [[CrossRef](#)] [[PubMed](#)]
  43. . Okusha, Y.; Hirai, Y.; Maezawa, H.; Hisadome, K.; Inoue, N.; Yamazaki, Y.; Funahashi, M. Effects of intraperitoneally administered L-histidine on food intake, taste, and visceral sensation in rats. *J. Physiol. Sci.* **2017**, 67, 467–474. [[CrossRef](#)]
  44. A. H. Abd Al-Ameer, " Metal Complexes of Mixed Ligands (Quinolone Antibiotics and  $\alpha$ -Aminonitrile Derivatives) Their Applications: An Update with Fe(III), Co (II) and Ni (II) Ions and Study the Biological Activity "*Journal of Global Pharma Technology*, Vol. 11, Issue 09 (Suppl.), pp. 515-524, 2019
  45. Vaziri, P.; Dang, K.; Anderson, G.H. Evidence for histamine involvement in the effect of histidine loads on food and water intake in rats. *J. Nutr.* **1997**, 127, 1519–1526. [[CrossRef](#)]
  46. Yoshimatsu, H.; Chiba, S.; Tajima, D.; Akehi, Y.; Sakata, T. Histidine suppresses food intake through its conversion into neuronal histamine. *Exp. Biol. Med. (Maywood)* **2002**, 227, 63–68. [[CrossRef](#)]
  47. Schmidt J.A., Rinaldi S., Scalbert A., Ferrari P., Achaintre D., Gunter M.J., Appleby P.N., Key T.J., Travis R.C. Plasma concentrations and intakes of amino acids in male meat-eaters, fish-eaters, vegetarians and vegans: A cross-sectional analysis in the EPIC-Oxford cohort. *Eur. J. Clin. Nutr.* 2016;70:306–312. doi: 10.1038/ejcn.2015.144. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
  48. Iwasaki M., Ishihara J., Takachi R., Todoriki H., Yamamoto H., Miyano H., Yamaji T., Tsugane S. Validity of a self-administered food-frequency questionnaire for assessing amino acid intake in japan: Comparison with intake from 4-day weighed dietary records and plasma levels. *J. Epidemiol.* 2016;26:36–44. doi: 10.2188/jea.JE20150044. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
  49. Steele B.F., Le Bovit C.B. Leucine and histidine tolerance in the human. *J. Nutr.* 1951;45:235–244. doi: 10.1093/jn/45.2.235. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
  50. Block W.D., Westhoff M.H., Steele B.F. Histidine metabolism in the human adult: Histidine blood tolerance, and the effect of continued free L-histidine ingestion on the concentration of imidazole compounds in blood and urine. *J. Nutr.* 1967;91:189–194. doi: 10.1093/jn/91.2.189. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
  51. Henkin R.I. New aspects in the control of food intake and appetite. *Ann. N. Y. Acad. Sci.* 1977;300:321–334. doi: 10.1111/j.1749-6632.1977.tb19332.x. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
  52. Okubo H., Sasaki S. Histidine intake may negatively correlate with energy intake in human: A cross-sectional study in Japanese female students aged 18 years. *J. Nutr. Sci. Vitaminol. (Tokyo)* 2005;51:329–334. doi: 10.3177/jnsv.51.329. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
  53. Nakajima S., Hamada M., Tsuchiya T., Okuda H. Suppression of food intake by histidine-rich protein under low energy intake. *Nippon Eiyu Shokuryo Gakkaishi J. Jpn. Soc. Nutr. Food Sci.* 2000;53:207–214. doi:

54. Schechter P.J., Prakash N.J. Failure of oral L-histidine to influence appetite or affect zinc metabolism in man: A double-blind study. *Am. J. Clin.Nutr.* 1979;32:1011–1014. doi: 10.1093/ajcn/32.5.1011. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
55. Geliebter A.A., Hashim S.A., Van Itallie T.B. Oral L-histidine fails to reduce taste and smell acuity but induces anorexia and urinary zinc excretion. *Am. J. Clin. Nutr.* 1981;34:119–120. doi: 10.1093/ajcn/34.1.119. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
56. Sasahara I., Fujimura N., Nozawa Y., Furuhashi Y., Sato H. The effect of histidine on mental fatigue and cognitive performance in subjects with high fatigue and sleep disruption scores. *Physiol. Behav.* 2015;147:238–244. doi: 10.1016/j.physbeh.2015.04.042. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
57. Layrisse M., Martínez-Torres C., Leets I., Taylor P., Ramírez J. Effect of histidine, cysteine, glutathione or beef on iron absorption in humans. *J. Nutr.* 1984;114:217–223. doi: 10.1093/jn/114.1.217. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
58. Schölmerich J., Freudemann A., Köttgen E., Wietholtz H., Steiert B., Löhle E., Häussinger D., Gerok W. Bioavailability of zinc from zinc-histidine complexes. I. Comparison with zinc sulfate in healthy men. *Am. J. Clin. Nutr.* 1987;45:1480–1486. doi: 10.1093/ajcn/45.6.1480. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
59. Yoganathan S., Sudhakar S.V., Arunachal G., Thomas M., Subramanian A., George R., Danda S. Menkes disease and response to copper histidine: An Indian case series. *Ann. Indian Acad. Neurol.* 2017;20:62. doi: 10.4103/0972-2327.199907. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
60. Jain P., Kannan L., Chakrabarty B., Kumar A., Gupta N., Kabra M., Gulati S. Menkes disease – An important cause of early onset refractory seizures. *J. Pediatr. Neurosci.* 2014;9:11–16. doi: 10.4103/1817-1745.131471. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
61. Christodoulou J., Danks D.M., Sarkar B., Baerlocher K.E., Casey R., Horn N., Tümer Z., Clarke J.T. Early treatment of Menkes disease with parenteral copper-histidine: Long-term follow-up of four treated patients. *Am. J. Med. Genet.* 1998;76:154–164. doi: 10.1002/(SICI)1096-8628(19980305)76:2<154::AID-AJMG9>3.0.CO;2-T. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
62. Snedeker S.M., Greger J.L. Metabolism of Zinc, Copper and Iron as Affected by Dietary Protein, Cysteine and Histidine. *J. Nutr.* 1983;113:644–652. doi: 10.1093/jn/113.3.644. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
63. Van Campen D., Gross E. Effect of Histidine and Certain Other Amino Acids on the Absorption of Iron-59 by Rats. *J. Nutr.* 1969;99:68–74. doi: 10.1093/jn/99.1.68. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
64. Darwish H.M., Cheney J.C., Schmitt R.C., Ettinger M.J. Mobilization of copper(II) from plasma components and mechanisms of hepatic copper transport. *Am. J. Physiol.* 1984;246:G72–G79. doi: 10.1152/ajpgi.1984.246.1.G72. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
65. Hartter D.E., Barnea A. Brain tissue accumulates 67copper by two ligand-dependent saturable processes. A high affinity, low capacity and a low affinity, high capacity process. *J. Biol. Chem.* 1988;263:799–805. [[PubMed](#)] [[Google Scholar](#)]
66. Mas A., Sarkar B. Uptake of 67Cu by isolated human trophoblast cells. *Biochim. Biophys. Acta BBA-Mol. Cell Res.* 1992;1135:123–128. doi: 10.1016/0167-4889(92)90127-W. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

67. McArdle H.J., Guthrie J.R., Ackland M.L., Danks D.M. Albumin has no role in the uptake of copper by human fibroblasts. *J. Inorg. Biochem.* 1987;31:123–131. doi: 10.1016/0162-0134(87)80057-0. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
68. Freeman R.M., Taylor P.R. Influence of histidine administration on zinc metabolism in the rat. *Am. J. Clin. Nutr.* 1977;30:523–527. doi: 10.1093/ajcn/30.4.523. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
69. Buxani-Rice S., Ueda F., Bradbury M.W.B. Transport of Zinc-65 at the Blood-Brain Barrier During Short Cerebrovascular Perfusion in the Rat: Its Enhancement by Histidine. *J. Neurochem.* 2002;62:665–672. doi: 10.1046/j.1471-4159.1994.62020665.x. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
70. Wensink J., Van den Hamer C.J. Effect of excess dietary histidine on rate of turnover of <sup>65</sup>Zn in brain of rat. *Biol. Trace Elem. Res.* 1988;16:137–150. doi: 10.1007/BF02797098. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
71. Van Wouwe J.P., Hoogenkamp S., Van den Hamer C.J. A histidine supplement and regulation of the zinc status in Swiss random mice. *Biol. Trace Elem. Res.* 1990;24:207–216. doi: 10.1007/BF02917208. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
72. Frezza C. Histidine metabolism boosts cancer therapy. *Nature.* 2018;559:484–485. doi: 10.1038/d41586-018-05573-4. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
73. Kanarek N., Keys H.R., Cantor J.R., Lewis C.A., Chan S.H., Kunchok T., Abu-Remaileh M., Freinkman E., Schweitzer L.D., Sabatini D.M. Histidine catabolism is a major determinant of methotrexate sensitivity. *Nature.* 2018;559:632–636. doi: 10.1038/s41586-018-0316-7. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
74. Ikezaki S., Nishikawa A., Furukawa F., Enami T., Mitsui M., Tanakamaru Z., Kim H.C., Lee I.S., Imazawa T., Takahashi M. Long-term toxicity/carcinogenicity study of L-histidine monohydrochloride in F344 rats. *Food Chem. Toxicol. Int. J. Publ. Br. Ind. Biol. Res. Assoc.* 1996;34:687–691. doi: 10.1016/0278-6915(96)00033-6. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
75. Gullino P., Winitz M., Birnbaum S.M., Cornfield J., Otey M.C., Greenstein J.P. Studies on the metabolism of amino acids and related compounds in vivo. I. Toxicity of essential amino acids, individually and in mixtures, and the protective effect of l-arginine. *Arch. Biochem. Biophys.* 1956;64:319–332. doi: 10.1016/0003-9861(56)90276-4. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
76. Kanarek N., Keys H.R., Cantor J.R., Lewis C.A., Chan S.H., Kunchok T., Abu-Remaileh M., Freinkman E., Schweitzer L.D., Sabatini D.M. Histidine catabolism is a major determinant of methotrexate sensitivity. *Nature.* 2018;559:632–636. doi: 10.1038/s41586-018-0316-7. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
77. Darwish H.M., Cheney J.C., Schmitt R.C., Ettinger M.J. Mobilization of copper(II) from plasma components and mechanisms of hepatic copper transport. *Am. J. Physiol.* 1984;246:G72–G79. doi: 10.1152/ajpgi.1984.246.1.G72. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
78. Hartter D.E., Barnea A. Brain tissue accumulates <sup>67</sup>copper by two ligand-dependent saturable processes. A high affinity, low capacity and a low affinity, high capacity process. *J. Biol. Chem.* 1988;263:799–805. [[PubMed](#)] [[Google Scholar](#)]
79. Ikezaki S., Nishikawa A., Furukawa F., Enami T., Mitsui M., Tanakamaru Z., Kim H.C., Lee I.S., Imazawa T., Takahashi M. Long-term toxicity/carcinogenicity study of L-histidine monohydrochloride in F344 rats. *Food Chem. Toxicol. Int. J. Publ. Br. Ind. Biol. Res. Assoc.* 1996;34:687–691. doi: 10.1016/0278-6915(96)00033-6. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
80. Gullino P., Winitz M., Birnbaum S.M., Cornfield J., Otey M.C., Greenstein J.P. Studies on the metabolism of amino acids and related compounds in vivo. I. Toxicity of essential

amino acids, individually and in mixtures, and the protective effect of l-arginine. Arch. Biochem. Biophys. 1956;64:319–332. doi: 10.1016/0003-9861(56)90276-4. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

81. Alderman G., Jarrige R. Feed Evaluation and Protein Requirement Systems for Ruminants. [(accessed on 2 March 2018)]; Available online: <https://publications.europa.eu/en/publication-detail/-/publication/72fd322e-2083-4a19-9105-758eda4faf9b/language-en>.
82. Toerien C.A., Trout D.R., Cant J.P. Nutritional stimulation of milk protein yield of cows is associated with changes in phosphorylation of mammary eukaryotic initiation factor 2 and ribosomal s6 kinase 1. J. Nutr. 2010;140:285–292. doi: 10.3945/jn.109.114033. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]