Effect of Insect Growth Regulators, Hydroprene and Methoprene Treatment on Oviposition of selected Three Mosquitoes

P. Rajasekar, M.Meeramaideen and M.Salahudeen

Assistant professor in P.G & Research Department of Zoology Jamal Mohamed college, Tiruchirappalli- 620020, Tamilnadu, (INDIA)

ABSTRACT

Mosquitoesaremedicallyandveterinaryimportantvectors, responsible for the transmission of many human and animal diseases, such as malaria, yellow fever, dengue and West Nile Fever. The present study was conducted under laboratory conditions and consisted to evaluate the ovipositionattractancy of Insect Growth Regulator(IGR)HydropreneandMethoprene,ontheovipositionofthreespeciesof mosquitoes, viz., *Culex quinquefasciatus*(Say), *Aedes aegypti*(L.) and *Anophelesstephensi*(Liston).

Keywords: Hydroprene, Methoprene, ovipositionattractancy, *Culex quinquefasciatus*,

Aedes aegyptiandAnophelesstephensi.

Corresponding author – rajasekar275@gmail.com

INTRODUCTION

For many viral, bacterial and protozoans' diseases mosquitoes act as vector (Kalita*et al.*, 2013). In term of disease transmission and public health importance mosquito are considered as very important group of insects. Population of mosquito's increases exponentially that is major problem for many countries because mosquito spread the different diseases such as filarial, Japanese encephalitis, Lyme disease, Yellow fever, encephalitis, malaria, chikungunya, dengue, and epidemic poly-arthritis(Kamareddine.2012). In tropical and subtropical countries mosquito borne diseases are main problem (Kalita*et al.*, 2013).

Mosquito has approximately 3500 species and present in tropical and subtropical regions (Ghoshet al., 2011). Major genera of mosquitoes that act as vector for various diseases are *Culex* (Japanese encephalitis, west Nile, chikungunya, *Anopheles* (filariasis, malaria) and *Aedes* (chikungunya, dengue, Yellow fever) (Ghoshet al.,2012).

Major cause for the chikungunya and dengue is *Aedes aegypti*that act as vector for the disease and affect the 2.5 million people every year(Mendkiet al., 2015). Feletti, vivaxGrassi, protozoal parasites, *Plasmodium ovalestephens* and *Plasmodium falciparum welch* are the major cause of malaria that are spread by *Anopheles* mosquito (Gaddagutiet al., 2016). Most important reason for the increase of dengue fever are increased breeding places for the *Aedes* mosquitoes, less effective control of mosquito, more urbanization and enhanced growth of population (Soonwera.2015).

Annually worldwide the 200 million-450 million infections are caused by the Anopheles mosquito that leads towards 2.7 million deaths. In more than 100 rising countries it remains endemic disease (Koech and Mwangi, 2014). Virus of Japanese encephalitis occurs in the children with malnutrition and present in the areas that are linked with animal reservoirs particularly with pigs. On the other hand main cause for the encephalopathy is encephalitis (Yoon *et al.*, 2015). According to the BBC world service health program the mosquito is considered as the world's most dangerous animal (Mendki*et al.*, 2015).

Mosquito is vector and it causes severe diseases which can lead toward death so accurate action is necessary to get away from the disease as soon as possible (Gaddagutiet al., 2016). There are various methods that can be adopted to protect itself from the bite of mosquitoes.

Insect growth regulators (IGRs), the third generation insecticides, are diverse groups of chemical compounds that are highly active against larvae of mosquitoes and other insects. The IGRs in general have a good margin of safety to most non-target biota including invertebrates, fish, birds and other wildlife. They are also relatively safe to man and domestic animals. The IGR compounds do not induce quick mortality in the pre-imaginal stages treated and occur many days post treatment. This is indeed a desirable feature of a control agent because larvae of mosquitoes and other vectors are an important source of food for fish and wildlife. On account of these advantages of IGRs and the high level of activityagainst target species, it is likely that IGRs could play an important role in vector control programs in the future (Mulla, 1995). They are more specific for mosquitoes than conventional insecticides. The IGRs interfere with the hormonal mechanisms of target organisms and result in various kinds of morphological, anatomical and physiological abnormalities so that the target species does not reach the final stage of development (Amalrajet al., 1988a). There is no likelihood of resistancedevelopment

against these IGRs. A large number of IGRs, both juvenoids and chitin synthesis inhibitors, have been evaluated for the vector control but only very few of these are found effective and commercially feasible, e.g., diflubenzuron, methoprene, fenoxycarb (Tyagi *et al.*, 1985, 1987; Amalraj*et al.*, 1988b, Vasuki, 1988). The IGRs have an added advantage of being used at a relatively very low dose compared to the conventional insecticides. Ecdysone agonists are hormonally active insect growth regulators that disrupt development of larvae and are found to be active against *Ae. aegypti*, *An. gambiae*, and *Cx.quinquefasciatus*(Beckage*et al.*, 2004). The present work has beendesigned to study the ovipositionattractancyofIGR, HydropreneandMethopreneagainstthemosquitoes*Cx. quinquefaseiatus*, *Ae. aegypti*and*An.stephensi*.

MATERIALS AND METHODS

Test insects

Cx. quinquefaseiatus, Ae. aegyptiand An. stephensimosquitoes were obtained from a stock colonybeingmaintainedintheinsectaryat27±1&C

and 75±5% relative humidity at laboratory, Department of Zoology, Jamal Mohamed College, Trichirappalli. 10% sucrose was provided to females. Female mosquitoes were fedor rabbit blood for 4-5 days. Fivedays after blood feeding, the gravid female mosquitoes were used for bio assay experiments.

Test chemicals

IGRcompoundsnamelyHydroprenechemically known as Ethyl (2*E*, 4E)-3, 7, 11-Trimethyldodeca-2, 4-dienoateureawasreceivedasgratis(15%ECformulation Malwa Colony, Patiala, Punjab.Methoprene20% ECchemicallyknownasisopropylPropan-2-yl (2*E*,4*E*)-11-methoxy-3,7,11-trimethyldodeca-2,4-dienoate.Supplied byChemiInternational,LokmanyaNagar,Mumbai,India.

Bioassay

Ovipositionbehaviour tests were carried out (Xueet al.,2001). Fifteengravidfemale mosquitoes(10daysold5days after blood feeding) were transferred to each mosquitocage (45x38x38cm).Concentrationsof0.0015, 0.0020and0.0025mg/Lweremadefromeach compoundsin100mlofwater.Twoenamelbowlsholding100mlofwaterwereplacedin Oppositecorners ofeachcage,onetreatedwiththetestmaterialand theotheronewasethanolcontrol.Thepositionsof thebowlswerealternatedbetweenthedifferentreplicates,soastonullifyanyeffectofpositiononovipo sition.Fivereplicatesforeachconcentrationwererun, withcagesplacedsidebysideforeachbioassay.The percenteffectiveattractancy(%EA)foreachovipositionconcentrationwascalculated (Kramer and Mulla.1979)andOvipositionActiveIndex(OAI)wereassessed (Graham-Bryce et al.,1987).

OAI - NT-NS

WhereNT-TotalnumberofeggsinthetreatedwaterandNS-Totalnumberofeggslaidinthecontrol water.

RESULTS

TheHydropreneovipositional responses of the normal gravid females of the three vector mosquito species to Novaluron treated water medium are presented in Table1. The results indicate that the number of eggs laid by the females was higher in control than in treated water. The total number of eggs laid in the treated water was also found to vary at different dosages. The OAI values calculated from the standard formula revealed that this IGR compound has repelling activity at higherdosages.

On comparing the OAI values of the three species Cx. Quinquefasciatus (-0.40), Ae. aegypti(-0.32) and An. stephensi(-0.30) showed considerable negative response at

the dose 0.010 mg/l. At the dose of 0.020 mg/l *Cx. quinquefasciatus*(-0.52), *Ae.aegypti*(-0.45) and *An. stephensi*(-0.55), showed a higher negativeresponse.

The ovipositional responses of the normal gravid females of the three vector mosquito species to Methoprenetreated water medium are presented in Table 2. The results indicate that the number of eggs laid by the females was higher in control than in treated water. The total number of eggs laid in the treated water was also found to vary at different dosages. The OAI values calculated form the standard formula revealed that this IGR compound has repelling activity at higher dosages.

On comparing the OAI values of the three species *Cx. Quinquefasciatus* (- 0.37), *Ae.aegypti*(-0.35) and *An. stephensi*(- 0.31)showed considerable negative response at the dose of 0.010 mg/l. At the dose of 0.020 mg/l. *Cx. quinquefasciatus*(-0.55), *Ae.* aegypti (-0.50) and *An. stephensi*(-0.65), showed higher negative response. On comparision of ovipositional responses of three vector mosquitoes to two IGRs compounds, Methopreneexhibited more ovipositional negative responses.

Table 1 Oviposition response of mosquitoes to Hydroprenetreated water

	Cx. quinquefasciatus			Ae. aegypti			An. stephensi		
Concentr ation (mg/l)	No. of egg rafts			No. of egg rafts			No. of egg rafts		
	Treate d	Control	O AI	Treate d	Control	O AI	Treate d	Control	O AI
0.015	45.66 ±1.21	101.16 ±3.97	0.40	52.0±2.52	95.83±1.4 7	0.32	39.33±1.8 7	93.83±2.13	- 0.30
0.020	41.16 ±1.16	100.16 ±2.92	0.43	48.66±1.2 1	95.55±1.5 1	- 0.40	36.83±1.1 6	94.5±1.04	- 0.38
0.025	37.66 ±1.03	102.21 ±3.04	0.52	45.66±1.2	94.32±1.2 8	0.45	33.53±1.9 4	94.16±1.63	- 0.55

OAI - Oviposition Active Index

Table 2 Oviposition response of mosquitoes to Methoprene treated water

Concentr ation (mg/l)	Cx. quinquefasciatus			Ae	e. aegypti		An. stephensi		
	No. of egg rafts			No. of egg rafts			No. of egg rafts		
	Treate d	Control	O AI	Treate d	Control	O AI	Treate d	Control	O AI
0.015	92.16±3 .04	207.53± 6.71	- 0.3 7	105.5±4 .03	205.71± 3.67	0.3	98.14±2 .44	203.83± 3.43	0.3
0.020	81.33±3 .65	210.13± 5.94	- 0.4 4	90.16±3 .94	208.16± 4.08	- 0.3 9	81.83±2 .94	208.56± 4.32	0.3
0.025	70.5±2. 86	212.64± 6.48	- 0.5 5	79.5±4. 08	209.7±4. 32	- 0.5 0	54.83±3 .71	216.16± 4.07	- 0.6 5

DISCUSSION

Mosquitoes are the most important single group of insects in terms of public health importance, which transmit a number of diseases, such as malaria, filariasis, dengue, Japanese encephalitis etc., causing millions of deaths every year (Rahuman*et al.*, 2009). Human beings are compelled to fight against them using available technical ornaments. There was initial success in controlling vectors by using synthetic insecticides. Since 1900, the World Health Organization has warned about the possible emergence and reemergence of arthropod-borne disease due to combined human, biological, environmental and climatic factors (WHO, 1990).

One of the approaches for control of these mosquito-borne diseases in the interruption of disease transmission is by killing or preventing mosquitoes to bite human beings. Although numorous synthetic pesticides are presently available for vector control programmes operating in many areas of the world (Walker, 2000; Curtis and Davis, 2001), the intense and wide spread use of these products has caused concerns regarding their impact on both human and environmental health, and has led to a buildup of resistance in pest population (Tremblay, 1982; Schmutter, 1990).

The most effective way to combat with this mosquito infestation is the prevention of mosquito breeding through the use of larvicides, synthetic insecticides such as organophosphates have

been used as larvicide in several countries for the last 30 years (Chavasse and Yap, 1997). However, one major drawback with the use of these chemical insecticides is that they are non-selective and could be harmful to other organisms in the environment (Omena*et al.*,2007). The comparison of OAI values indicated that the IGRs compounds eliciting repelling activity. It is quite possible that this IGR acts more as an ovipositiondetterent inhibiting oviposition at higher concentrations in the oviposition medium than as an oviposition repellent that causes insects to make oriented movements away from the source as proposed by Rajasekar and Jebanesan, (2011).

The oviposition deterrent activity exhibited by this IGRs relatively higher when compared to that of few insecticides like Pyriproxyfen (0.3 or 0.4mg/cm²) against Ae. aegyptimosquitoes Chism and Apperson(2003).

Ganesan*et al.* (2006) have found *Ae. aegypti*females not responding to oviposition substrates treated with 100 ppm of certain C12-C18 fatty acids and their methyl esters identified from the eggs, synthetic pyrethroids like cypermethrin, fentalerate, decamethrin and permethrin have also shown repellency against *Cx. quiquefasciatus*, *Ae. aegypti* and *An. stephensi* laboratory oviposition experiments (Verma, 1986). However, another IGRs triflumuron has been reported to elicit increased oviposition by *Ae. aegypti* field and laboratory experiments (Ziechner and Peric, 1999).

The results of present study are interesting. The findings indicate the importance of traditional knowledge in science. The selected two IGR compounds Hydroprene and Methoprene were tested against the mosquito in laboratory condition for oviposition deterrent activity. On comparison of ovipositional responses of three vector mosquitoes to two IGRs compounds, Methopreneexhibited more ovipositional negative responses.

REFERENCES

- 1. Kalita B, Bora S, Sharma AK. Plant essential oils as mosquito repellent-a review. *International Journal of Research and Development in Pharmacy and Life Sciences*. 2013;3(4):741-747.
- 2. Kamareddine L. The biological control of the malaria vector. Toxins. 2012;4(3):748-767.
- 3. Ghosh A, Chowdhury N, Chandra G. Plant extracts as potential mosquito larvicides. Indian J Med Res. 2011; 135(46):581-598.
- 4. Ghosh A, Chowdhury N, Chandra G. Plant extracts as potential mosquito larvicides. Indian J Med Res. 2012; 135(23):581-598.
- 5. Mendki MJ, Singh AP, Tikar SN, Parashar BD, Veer V, Shukla SV et al. Repellent activity of N, N- diethylphenylacetamide (DEPA) with essential oils against Aedes aegypti, vector of dengue and chikungunya.International Journal of Mosquito Research. 2015;2(1):17-20.
- 6. Gaddaguti V, Rao TV, Rao AP. Potential mosquito repellent compounds of Ocimum species against 3N7H and 3Q8I of Anopheles gambiae.Biotech.2016; 6(11):1-8.
- 7. Soonwera M. Efficacy of essential oils from Citrus plants against mosquito vectors Aedes aegypti(Linn.) and Culex quinquefasciatus(Say). J Agri Tech. 2015; 11(2):669-681.
- 8. Koech PK, Mwangi RW.Synergistic repellent activity of plant essential oils against Aedes aegyption rabbit Skin.International Journal of Mosquito Research. 2014; 1(14):55-59
- 9. Belmadani, AATIKA BOUBEKKA KAHINA, and F. Mouhouche. "Insecticide activity of Daturainnoxia on migratory locust: Locustamigratoria (linné, 1758)(oedipodinae, acrididae)." Int J AgricSci Res 4.5 (2014): 1-8.
- 10. Yoon JK, Kim KC, Cho Y, Gwon YD, Cho HS, Heo Y. Comparison of repellency effect of mosquito repellents for DEET, citronella, and fennel oil.Journal of parasitology research.

2015;2015:1-6.

- 11. MendkiMJ,SinghAP,TikarSN,ParasharBD,VeerV,ShuklaSVetal.Repellentactivityof N, N-diethylphenylacetamide (DEPA) with essential oils against Aedes aegypti, vectorof dengue and chikungunya.International Journal of MosquitoResearch.2015;2(1):17-20.Hassan,F.A.I.Z.A.N.,DIWAKARSINGHDinesh,andB.I.D.Y.U.T.Purkait."BiochemicalcharacterizationofdetoxifyingenzymesinDDTresistantfieldisolatesofPhlebotomusargenitipe s in Bihar, India. "International Journal of Medicine and Pharmaceutical Sciences 5.2 (2015): 23-32.
- 12. MullaMS.The future of insect growth regulators in vector control.J. Am. Mosq. Control Assoc.1995; 11: 269-73.
- 13. Amalraj D, Vasuki V, Kalyanasundaram M, Tyagi BK Das PK. Laboratory
- 14. and field evaluation of three insect growth regulators against mosquito vectors. Indian J. Med.Res. 1988a; 87: 24-31.
- 15. Amalraj D, Vasuki V, Sadanandane C, Kalyanasundaram M, Tyagi BK, Das PK. Evaluation of two new juvenile hormone compounds against mosquito vectors. Indian J. Med. Res. 1988b; 87:19-23.
- 16. Tyagi BK, Kalyanasundaram M, Das PK, Somachary N. Evaluation of a new compound (VCRC/INS/A-23) with juvenile hormone activity against mosquito vectors. Indian J. Med. Res. 1985; 82:9-13.
- 17. Tyagi BK, Somachari N, Vasuki, V, Das PK. Evaluation of three formulations
- 18. of a chitin synthesis inhibitor (fenoxycarb) against mosquito vectors. Indian J. Med. Res. 1987;85:161-167.
- 19. Vasuki V. Role of insect growth regulators in vector control.Proc. 2ndSymp. Vector –borne Diseases, Trivandrum.1988;204-218.
- 20. Beckage NE, Marion KM, Walton WE, Wirth MC, Tan FF. Comparative
- 21. larvicidal toxicities of three ecdysone against on the mosquitoes Aedes aegypti, Culex quinquefasciatus, and Anopheles gambiae. Arch. Insect Biochem. Physiol. 2004;57: 111-22.
- 22. Mahgoub, MOSTAFA M., and NAGWA E. Azrag."Impact of Attitude and Knowledge of Insecticide Safety Household and Spray-Men on Gambiae Control Project Area Northern State, Sudan."International Journal of Zoology and Research (IJZR), 2018. 8 (1): 10.
- 23. Xue, R.D., Bernard, D.R and Ali, A. Laboratory and field evaluation of insect repellents as larvicides against the mosquito An. albimanus and Ae.albopictus. Med.
- 24. Vet. Entomol.2001;15: 374-380.
- 25. Kramer, L.W and Mulla, S.M.Oviposition attractants and repellents of mosquitoes: oviposition responses of Culexmosquito to organic infusions. Environ. Entomol. 1979; 8: 1111-1117.
- 26. I.J.Graham-Bryce, A.J.Eds, Burnth, Coaker, P.C.Jepson; Chemical Methods in Integrated Pest Management. 1987; 113-161.
- 27. Rahuman, A.A., Bagavan, A., Kamaraj, C., Saravanan, E., Zahir, A.A and Elango, G. Efficacy of larvicidal botanical extracts against Culex quinquefasciatusSay (Diptera: Culicidae).Parasitol.Res. 2009;104:1365–1372.
- 28. WHO. 1990. Report of the vector Biology and control, Geneva. Switzerland.3rd ED Walker, K. Cost-comparison of DDT and alternative insecticides for malaria control.Med.Vet. Entomol2000;14: 345–354.
- 29. Curtis, C.F and Davis, C.R. Present use of pesticides for vector and allergen control and future requirements. Medical and Veterinary Entomol. 2001;8: 231–235.
- 30. Tremblay, K.D. Rural-urban residence and concern with environmental quality: A replication and

extension. Rural Sociology.1982;43: 479-491.

- 31. Schmutter,H. Propertiesandpotentialofnaturalpesticidesfromtheneem tree, Azadirachtaindica.Annu.Rev. Entomol.1990; 35:271-297.
- 32. Chavasse, D.C. and Yap, H.H. 1997. Chemical methods for the control of vector and Pests of public health importance. WHO/CTD/WHOPES/97.2. WHO, Geneva, Switzerland Omena, M.C. de., Navarro, D.M.A.F., Paula, de. JE., Luna, J.S., Ferreira, de., Lima,
- 33. M.R and Sant'Ana, A.E.G. Larvicidal activities against Aedes aegyptiof some Brazilian medicinal plants.Bioresour.Technol.2007; 98: 2549- 2556.
- 34. Rajasekar, P and Jebanesan, A. Emergence inhibition effect of insect growth regulator Novaluron and Buprofezin against mosquitose (Diptera: Culicidae). J. Pest. 2011;35: 34-36.
- 35. Chism, B.D and Apperson, C.S. Horizontal transfer of the insect growth regulator pyriproxyfen to larval microcosms by gravid Aedes albopictusandOchlerotatustriseriatusmosquitoes in the laboratory.Med. Vet. Entomol.2003;17: 211-220.
- 36. Ganesan, K., Mendki, M.J., Suryanarayana, M.V.S., Prakash, S and Malhotra.R.C. Studies of Aedes aegypti(Diptera: Culicidae) ovipositional responses to newly identifiedsemiochemicals from conspecific eggs. Aust. J. Entomol. 2006;45: 75–80.
- 37. Verma, K.V.S.Deterrent effect of synthetic pyrethroids on the oviposition activity of mosquitoes.Curr.Sci. 1986; 55: 373-375.
- 38. Ziechner, B.C and Perich, M.J. Laboratory testing of a lethal ovitrap for Aedesaegypti. Med. Vet. Entomol1999;13:234-238.