

ARTIFICIAL PHOTO SYNTHESIS: BIOMIMIC TOOLS BEYOND NATURE

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ABSTRACT

In our surrounding the cellular of casing the molecular tools the chloroplasts probably playing the most important tool for plants. In the last decades, genomics and genetic and knowledge have acknowledged progress in the detection and functioning of chloroplasts. "Chloroplasts", the lime difference structure of 'plastids', there is an important part of photosynthesis and supplementary vital plant functions. In nature, photophosphorylation comes into existence in chloroplasts becoming the supreme prototype of ATP production. In presence of light, Photosystem II can fragment H₂O into protons, electrons, and oxygen. In the production of ATP, the process can create a proton gradient for ATPase. Therefore, an affectedly planned chloroplast for Photosystem II driven, Adenosine triphosphate (ATP) synthesis is comprehended. Indeed, the example of unlike chloroplast Anglo Saxon (Au)/chloroplast morph Titanium dioxide by original chloroplast nanostructure and alike purposeful structures are given. The "nanolayered", erections concerning the pattern of chloroplast which inherited within the chloroplast morph Titanium dioxide points into a huge response part and shown a fast 'Photo Induced Electron Transfer' as well as the fast reaction of co-catalyst Aurum (Au) nanoparticles which efforts as response endorse Photo Encouraged Charge detachment plus upgrade the whole photo-catalytic movement which helps in a chemical reaction. This theory delivers the inspiration for building efficient photo-catalysts, mimicking the photosynthesis course of equally desirable structures and functions. Our knowledge that the chloroplast process which notably suspends in a mixture of liquid/organic mix-ups which is fluid at sub-zero temperature with good maintenance of the capability to lessen synthetic of e acceptors. The working and their relations are proceeding quickly and making the organelle an important bull for structures biology research in plants.

Keywords

Photosynthesis, Genomics, Genetic, Nanostructure, Photocatalyst, sub-zero temperature.

Introduction

The light-harvesting technology of plants with enzymes from nine dissimilar organisms, scientists statementconstruction an artificial chloroplast that operates external of cells to harvest sunlight and custom the energy to change carbon dioxide (CO_2) into energy-rich molecules.

The artificial chloroplasts are accomplished of binding and changing the greenhouse gas carbon dioxide using light energy. Many years ago, Plantscould harness CO_2 from the air using solar energy. The Marburg research team led by director Tobias Erb has succeeded in completely created a platform for the automated structure of cell-sized photosynthetically active sections, "artificial chloroplasts" that can grasp the energy and then change the greenhouse gas, CO_2 with light.

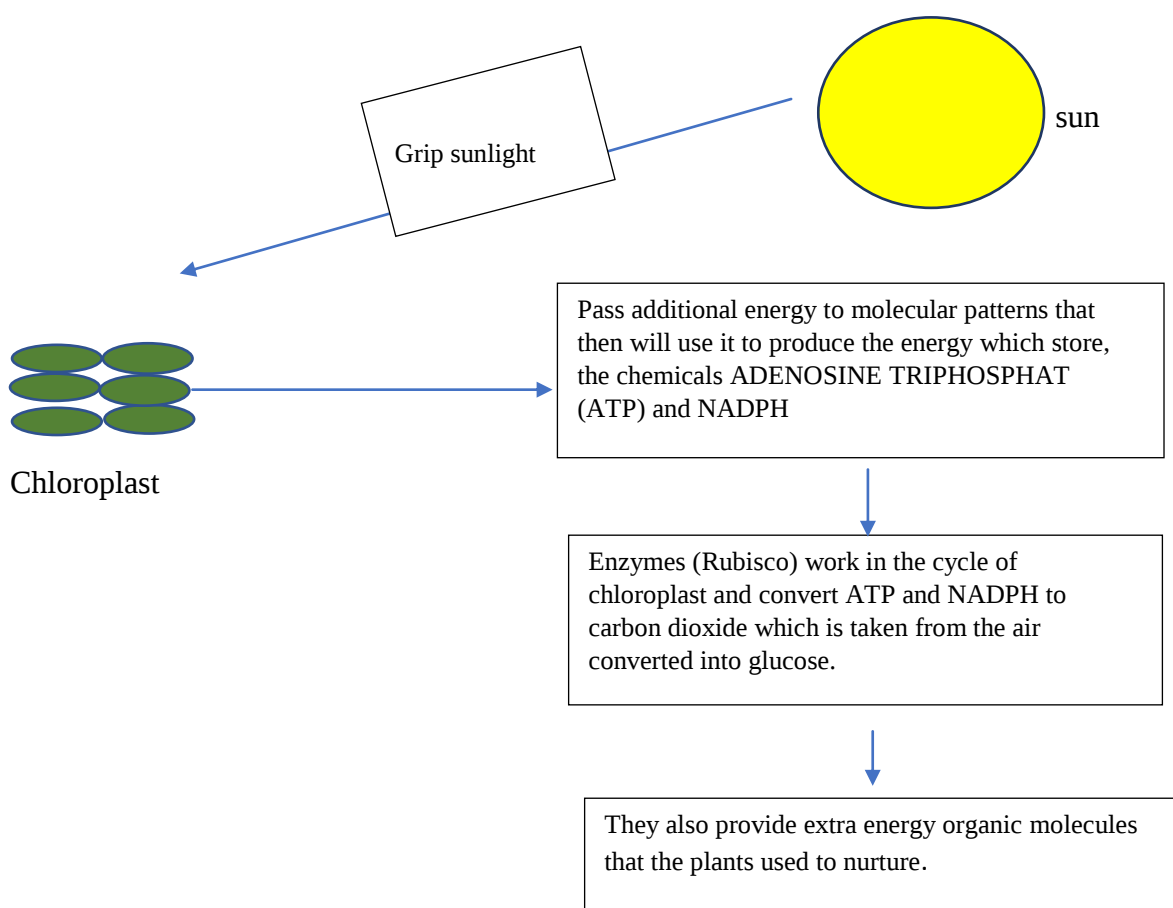


Figure 1: Process of artificial chloroplast

Chloroplast biology: Chloroplast study in The Genomic Phase-

Plant cells are consisting of organelles named plastids. The endosymbiotic is free of alive cyanobacterial progenitor plastids, who are required to end the progression period, losing the immense majority of their genes. Further proplastids can advance convert into a different

form of a pigment like emerald colour chloroplasts, yellow or red chromoplasts, or additional alternates skilled for the storage of lipids, proteins, and starch. The photosynthetic process, that is characterized by a lively chloroplast by high rates of translation and transcription, allowing them to synthesize on the great quantities of the enzyme (Ribulose biphosphate carboxylase) Rubisco and quick renewal of electron-transfer elements, features essential for effective and efficient photosynthetic carbon dioxide fixation. Other than the photosynthesis process, chloroplasts, transmit out different vital plant functions, like the synthesis of AA(amino acids), Lipids, and Fatty acids plant hormones, secondary metabolites, nucleotides, and vitaminsupplements. Scientific expansions of the genomics- *Arabidopsis thaliana*, with the genome sequencing, which have lately stirred the incentive of projects intended to have systematically identified functions of chloroplast' proteins, and their indoctrination Genes, and educating us about the evolutionary reasonings. The objecthighpoints new conclusions, concentrating on platforms for new chloroplast mutants, systematic reverse genetics, transcriptomics, bioinformatics, and proteomics', in an *Arabidopsis thaliana*, and supplementary types of plant.

Nuclear-encoded acquires mainly the chloroplast proteins and, with the exclusion of the external packed proteins, necessitate Nterminal, pre sequencing naming it as (CTPs)Chloroplast Transit Peptides, to bull them towards the chloroplast. In the range of 2100 - 3600 different types of proteins, which are projected to be in the '*Arabidopsis*', chloroplast^{1,2}.

Many studies, done the analyses of phylogenetic, that indicate the whole *Arabidopsis* genome/ samples of it, provision of the decisionamongst the range of 400 to 2200, *Arabidopsis* genes which mayhap of 'cyanobacterial' origin^{2,3}. The furthemost detailed study on the subject of Cyanobacterial heritage in the *Arabidopsis* nuclear genome is linked with genomes of trio cyanobacterial species. like prokaryotes, yeast, and 16 others⁴.

Inverse genetic examination of plastome genes-

Open reading frames (ORFs) prearranged by plastome DNA that belong to 2maincategories:

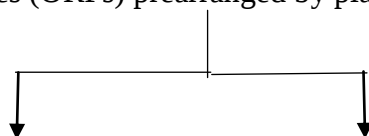


Table 1.Essential genes and Photosynthesis- linked genes.

1) Essential Genes	2) Photosynthesis - linked genes
Genes essential is used for the appearance of the organelles'particularsystem of genetic (indoctriation tRNAs, rRNAs,proteins, ribosomal, RNA polymerase subunits).	The chloroplast genome has ageneportion that is small, whichisrequired for the photosynthesis process, whereas the nuclear genome encodes others. First of all,the genetics of photosynthesis reports, the basic genes for the apo-proteins of macro-molecular enzymes and complexes that are essential in the process.

Comparatively, some hypothetical functions of plastid i.e., ORFs are not known. In chloroplast, efficient homologous recombination permits wild-type *yfc* genes to be changed through knock-out alleles, and in homo-transplastomic plants into their phenotypes deliberate^{5,8,9}.

Ahead genetics-awnings of mutant in a novel quantity and quality.

Currently, forward genetics is established as an excessive ideal of consideration in plant chloroplast research and studies, retaining phenotypic awnings for an extensive range of chloroplast progression. The first orderly studies founded on different chlorophyll (fluorescence) which features are considered as the high Chlorophyll Fluorescence Phenotype (hcf), which is observed as fluorescence red into Ultraviolet light in a plant. The hcf mutants are familiar in *C. reinhardtii*, *Arabidopsis*, and maize^{10,11,12}. In the maximum appearance, photosynthesis is affected severely, regularly subsequent in the lethality of a seedling. HCF genes are intricate in vital chloroplast roles, like 'thylakoid biogenesis'^{13,14,15}, 'protein translocation'¹⁶ 'mRNA processing'^{17,18} and 'translation'¹⁹. Parameter of non-photochemical quenching also called as NPQ, that imitates the energy to dissolve, as heat succeeding energization of the membrane of thylakoid due to lumen acidification /or, PSII quantum yield of real FII amount of the general efficacy of PSII response in the light center are supervised. 'NPQ' mutants are primarily present in *Chlamydomonas*²⁰, it is also recognized *Arabidopsis*, mutants that display variations in NPQ^{21,22}. The genes are protein scramble which involves in the cycle of xanthophyll in the process²¹. The genes encode the proteins which involved in the cycle of xanthophyll²¹, or such as PSII-S for sub-units of the photosynthetic (apparatus) kit²³ and also for the Rieske protein, of the 'cytochrome b6f complex'²⁴. The *Arabidopsis* mutants with dissimilar FII²⁵, disclose the flaws in proteins i.e., non-plastid protein and plastid protein with the PSI of subunit E²⁶, The 'Plastid Ribosomal Protein' (PRPL11)²⁷, and the 'Iron Transporter' (IRT1)²⁸. Of the independent, 12 mutant alleles linked with the vicissitudes in FII so far identified. The plastid protein 6 encoded and also 6 for proteins deprived of chloroplast, target it.

Table 2: The table shows the no. of mutant, cloned genes of chlorophyll fluorescence in *Chlamydomonas reinhardtii* chloroplast.

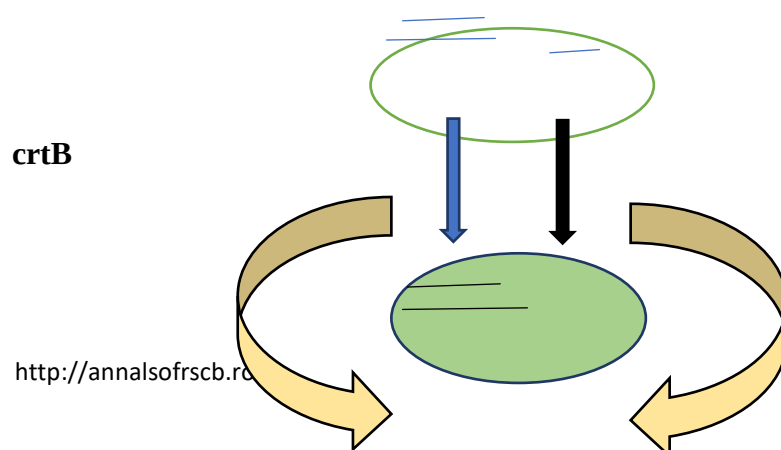
Phenotype	Mutants	No. of mutants	cloned Genes
'Chlorophyll fluorescence'			
'HCF'	'High Chlorophyll Fluorescence'	'67 ^a	'AtHCF136' ¹³ , 'AtHCF164' ¹⁴ , 'AtVIP1' ¹⁵ , 'AtHCF107' ¹⁸
'Several hundreds'		'b'	'ZmHCF106' ¹⁶ , 'ZmCRP1' ¹⁷ , 'ZmPRPS17' ¹⁹
'NPQ'	'Non-Photochemical Quenching'	'119 ^a	'AtZEAXANTHIN EPOXIDASE' ²¹ , 'AtVIOLAXANTHIN DEEPOXIDASE' ²¹
			'AtZEAXANTHIN EPOXIDASE' ²¹ , 'AtVIOLAXANTHIN'
			'DEEPOXIDASE' ²¹ , 'AtPsbS' ²³ , 'AtPetC' ²⁴
		'150 ^c	'CrNPQ5 (Lhcbm1)' ²⁹
'FII Effective Quantum yield of PSII'		'63 ^a	'AtPsaE1' ²⁶ , 'AtPrpl11' ²⁷ , 'AtIRT1' ²⁸

Early studies on chloroplasts are contingent mainly on the practice of biophysical and biochemical approaches. In the year 1980 and 1990, approaches that time was advanced for changing chloroplasts through homologous recombination, and for analytically disrupting nuclear genes by presenting T-DNAs or transposons. These advances decidedly enhanced the efficacy of approaches in genetic. To the studies of chloroplast functions. By the sequencing the of complete the genome and formation of high-throughput techniques for the analysis, chloroplast studies also arrived in the period of genomics. Functional genomic the research on proteomes, metabolomes, and transcriptomes—which immense the opportunities for the clarification of chloroplast roles which serve equally to describe the mutants and for classifying applicable loci that are for targeted mutagenesis.

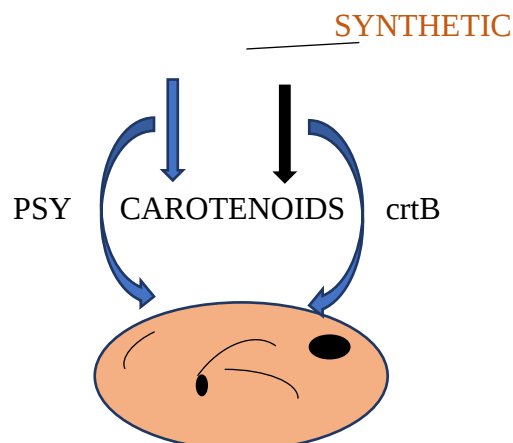
Artificial conversion of leaf chloroplasts addicted to carotenoid-rich plastids:

Plastids contain a collection of functionally and morphologically varied organelles in plants which accomplished of distinguishing form of plastid, additional in the response in the progression and environmental incentives^{30,31}. Alike, plastidial adaptations are vital to tolerate many processes in biological and show a huge back to the specialization of cell in dissimilar plant tissues. In the middle of the dissimilar types of plastid, the chromoplasts are present at a position in our creation and agriculture since their capacity to collect carotenoids of high levels, pigmented plant like isoprenoid delivers colour in the range of between yellow to red^{32,33,34}. Carotenoids like β -carotene, (provitamin A) are health endorsing supplements that cannot be manufactured by the animals but directly taken in their food, that are similarly added value complexes generally used in food, feed, cosmetics, and pharma productions as natural phytonutrients and pigments^{33,34,35}. Chromoplasts discriminate with some of the pre-existing plastids like proplastids. The Chloroplasts alter into chromoplasts through the expansion of several fruits and flowers, but only rare plant classes distinguish leaves from the chromoplasts^{30,31,32,33,34}. Some leaves contain colour from yellow to red that obtains as they grow (Example: when they are exposed to continuous darkness or autumn) because the carotenoids present in chloroplast become visible when chlorophylls damage. This process of senescence, however does not include the alteration of chloroplasts, into chromoplasts but into an entirely dissimilar kind of plastids termed as 'gerontoplasts'^{30,31}. The most prominent disparities throughout chloroplast $\rightarrow$ (into) chromoplast variation are the restructuring of the inner structures of plastid, composed with simultaneous harm of photosynthetic, overaccumulation and capability of pigments of carotenoid^{30-32,34,36,37}. The renovation of the inner plastid structures generates a high rate of metabolic capacity to sink but then it too endorses biosynthesis of carotenoid.

CHLOROPLAST



NATURAL CUESYSTEM



CHROMOPLAST

Figure 2: Synthetic conversion of leaf chloroplasts addicted to carotenoid-rich plastids.

The regulator of chromoplast difference seems as approaching for refining the dietary and strength's sake (i.e., benefits) of yields^{34,35,36,37,38}. The overall process is known to involve variations in gene expression (e.g., via retrograde signalling from plastids to the nucleus), hormonal regulation, protein quality control, and plastid protein import.

Congregation of 'ATPase and Photosystem II' for Light-Driven ATP Synthesis as unnatural chloroplast:

Through photophosphorylation and oxidative phosphorylation, ATP is produced by 'Proton Gradient' driven from adenosine triphosphate (ATP) synthase, FoF1-ATPase^{39,40,41}. Up to at the movement consideration is taken on an entity of 'ATPase based entities by re-formation of subcellular functional units, in artificial constructs, functioning as 'proteoliposomes'^{42,43}, 'polymersomes'⁴⁴ and polymer-based or protein or micro-capsules^{45,46,47}. A non-natural proton gradient in those structures in artificial proton gradient has been created by direct Pump' of pH⁴⁸, 'Enzyme Induced Response'⁴⁹ and 'Light-induced proton transfer', through Trans-membrane Photo-active Bacteriorhodopsin^{50,51,52} 'photosensitive artificial porphyrin antennas' or regulated 'chromatophores',^{53,54} Energy supply provided by the system of light-driven, by the simple remote regulator of light off/on.

With the ability of water to oxidize is, the only protein compound is photosystem II (PSII), is driving Photo-phosphorylation in plants. Taking close infrared light and visible, PSII initiates H₂O oxidizing into electrons, oxygen and protons at the cluster of Mn₄CaO₅.^{55,56} Protons unconfined and alteration by the connector with the Electron Transport Chain, (ETC) subsidize to the Proton gradient, for 'ATPase' to substitute and catalyse the combination of Adenosine triphosphate.^{57,58} Photoactive molecules related with others, can be immobilized PSII on practically adapted exteriors / captured in reticulate 'sol-gel cavities' by noncovalent OR covalent relations^{59,60,61,62,63,64,65}.

Light Driven ATP Synthesis Illustration:

Enthused by our nature, it is studied that in chloroplast photosynthetic chain is present, a concept is has formed a novel 'FoF1-ATPase' proteoliposome coated PS-II microspheres with

a Core@shell, construction via molecular assemblage to emulate the illumination toward the (ATP) adenosine tri phosphate alteration procedure happening in the sheath of the thylakoid.

The additional portion, Hydrogel similar Photosystem II based microparticles are arranged and organized through coprecipitate of (Bovine Serum Albumin) BSA.

(Calcium Carbonate) CaCO_3 and Photosystem II, followed by (Glutaric Dialdehyde) GA.

Cross-linking end ensuing exclusion of the (Calcium Carbonate) CaCO_3

Then 'FoF1 -ATPase' Proteo-liposomes are glazed arranged of the microsphere, exterior that revealed to figure a general chloroplast like, illumination (light) induced non-natural preparation of photophosphorylation. Beneath irradiation of red light, Photosystem II captured in the hydrogen similar solid-Microsphere separations H_2O into protons, which prompts successively the turning catalysis of 'FoF1-ATPase' of Proteo-liposomes on the carapace. By decoupling form of it the natural Photo-phosphorylation pathway, slightly this method of organelles sized Bio-reactor-likely forms this to evade PS I and 'Cytochrome b6f'. This categorization and combination approach can check away for original co-assembly, a unit of multi-functional and representing multipart bio-chemical process.

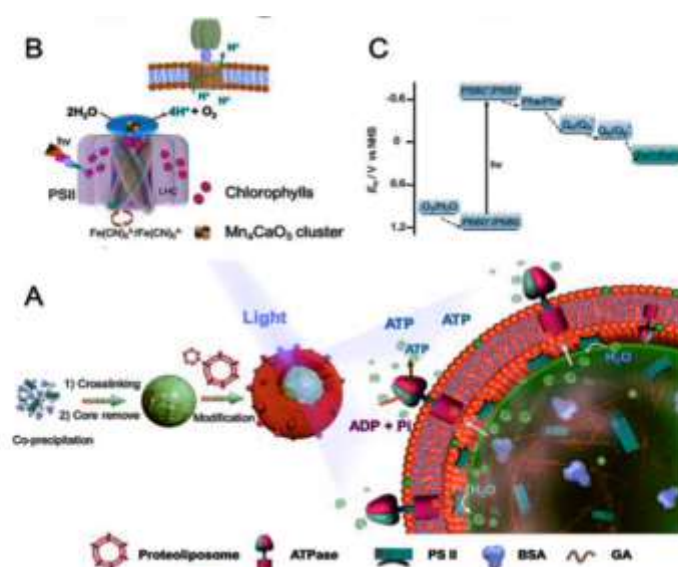


Figure 3:

The Picture of illumination of (Light) Driven adenosine triphosphate (ATP) Combination: (A) the form of 'CFoF1-ATPase' (B) ionDeparture, also shoe transport of electrons, breakdown of H₂O, and proton-flow in the system of microsphere-based on PS II; (C) Integrated System, the scheme of Redox Potential which process as Electron transfertechnique.

Artificial Chloroplast prototype, with chloroplast purpose and erection:

One of the renewables and important energy, Solar Energy, a cleansed form of energy resources, shows an important part in resolving environmental difficulties and global energy. Solar form to chemical form of energy modification, an economical and efficient way for utilizing the solar energy⁶⁶. Distinctive representative of a photosynthesis, in billions of years nature has employed the delivery of energy for the whole environment, as well as to continue level of normal Oxygen and Carbon di oxide in the aerosphere⁶⁷. The chloroplast, is the place of 'photosynthesis', in green plants. whose 'Nanolayered' erections elaborate and ordered extremely useful mechanisms which results in utilization of efficient light which is visible, remarkable light harvesting, rapid Photoinduced ion separation, highly efficient photosynthetic activity and thus high energy transfer⁶⁸. However, the rate of various photosynthetic organisms varies with seasons, and it is also noted. Therefore, on annual basis photosynthesis is remarkably inefficient, with energy alteration productivity⁶⁹.

Chloroplast-morph-Titanium dioxide chloroplast construction and achieves Nitrogen and Phosphorus self-doping:

Enthused by innate chloroplast, for an example of unlike chloroplast, gold (Au)/chloroplast-morph-Titanium dioxide inorganic non-natural system of photosynthesis with nano layered structure of chloroplasts and also features of functional analogous. Chloroplast-morph-titanium dioxide was ready with normal chloroplast as the bio template. Chloroplast-morph-titanium dioxide gets the nanolayered erections of the template of chloroplast, it can show and leads to a reaction, which is a large reaction area and also show speedy photoinduced electron transfer;⁷⁰ gold (Au, element 79) nano-particles the cocatalyst, function as response middle point to endorse Photoinduced ion departure. Also, the improvement of overall Photo-catalytic action in H₂ creation; Nitrogen, Phosphorus self-doped from bio-template can upsurge observable light or sun shine absorption, alike to the pigments of projection. With the system of artificial chloroplast, it was found that the test samples with rapid photoinduced electron transmission and Various improved Photo-catalytic actions in degradation of pigment and production of H₂.

For developing, efforts have been dedicated well-organized non-natural system of photosynthesis, by its nature repeating the method of natural photosynthesis with correctly considered 'Supra-molecular' representing of photosynthesis' donor photosensitizer structure⁷¹, cyanobacteria or active thylakoids entrapping into a 3-dimensional network of silica to build a steady photosynthetic bio-reactor⁷², chlorosomes depositing onto Titanium dioxide to forge a bio-mimetic light harvesting tool⁷³, and so⁷⁴ Water, solar splitting into H and O with inert semi-conductor resource that is also observe as an main part of non-natural photosynthesis structure⁷⁵. Under UV light as the high efficiency of photo-catalyst, Titanium Dioxide (TiO₂)

has been applied broadly in biological impurity deprivation⁷⁶, intense of H^+ ⁷⁷, Dye sensitized, solar cells⁷⁸, and, self-cleansed⁷⁹. To attest to the truth or validity of conformation of the composition modifying of titanium dioxide, like Nonmetal components doping⁸⁰, doping of metal ion⁸¹, and loading of noble metal⁸², might boost the Photo-catalytic action. Furthermore, several classified⁸³ or bring together, designs based on nano-tube⁸⁴, nano-wire⁸⁵, etc. have discovered more efficiency Photo-catalytic action. Subsequently complex microstructure directly synthesis and that work is not a simple, pattern-based approaches are employed often⁸⁶. Related to mutual patterns, biotemplates moreover provides non-metallic compounds for self-doping⁸⁷ but also attractive classified constructions⁸⁸. As yet, it is an immense trial still bring to together effective Light-harvesting, and rapid ion separation, high transfer of electron and well erections into an 'integrated system. The consideration to the 'Highly Effective Processes of solution, for a nature. Chloroplasts' functional features and special structures progress their efficiency of photosynthetic. The nano-layered erection of the membranes of thylakoid which delivers a very great area of surface, to burden which is, numerous pigments of photosynthetic which has been captured by illumination and consequently develops the collecting of sunshine efficacy. The projection tints can be eager late taking red and blue light⁸⁹, which accounts largely for a part in the solar, (range) spectrum. Then the transfer of excitement energy, can be rapidly form the pigments to the response centres, wherever the departure of charge arises and the next replays is done⁹⁰. A blue-print which is provided by our nature, for scheming an effective 'false Photo-Synthesis system'. The collective chloroplasts,' biologically characterised and inert semi-conductor Titanium dioxide's photo-catalytic possessions composed, and future the example of "artificial chloroplast".

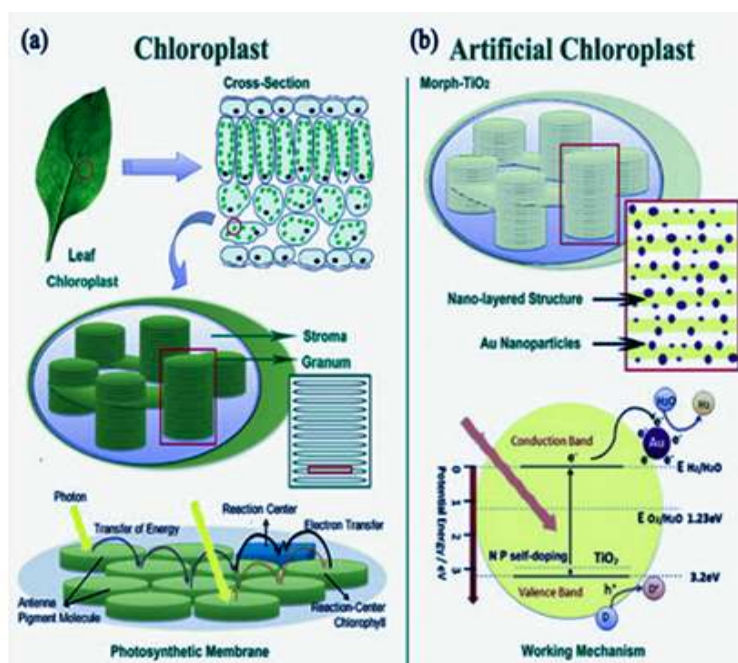


Figure 4: Artificial chloroplast: Au/chloroplast-morph-TiO₂ with fast electron transfer and enhanced photocatalytic activity.

‘Artificial chloroplast’ further creates gold (Au, element no: 79) chloroplast morph (Titanium dioxide) TiO_2 . In-organic unnatural photosynthesis’ structure as a precursor. The precursor that can benefit to progress extra separation of H_2O resources, with congenital usual chloroplast’s nanolayered construction and analogous efficient features to attain higher photo-catalytic action. A recent approach was done and also verified and aimed, to represent the process in natural way, to do the formation and formation of an, ‘Artificial Photosynthesis System’ for effective Renewable Resources.

Artificial Chloroplast Electron Acceptor Reduction at Sub- Zero Temperatures by Chloroplast Deferred in Liquid Media:

For the study of bio-chemical reaction one of the most powerful search is of, the lowering outcome of the temperature. In the interest of specific instance responses, like (ETC) Electron Transport Chain. The Temperature at the range of ‘-60 & 0 °C’, not only they detect prime photo-chemical response of reactions, but also the second responses of the reaction, the level that must berise as the condition is increased. In liquid media such temperatures can be carried out, Also in the frozen state. Newly research shows the examination of bio-chemical responses in liquid-medium at Subzero condition of the temperature, was delayed absence of information on the physicochemical assets on liquid or biological combinations necessarily i.e., solvent. In the laboratory, such data has been obtained now^{91,92} together with practice with a variation in enzymes isolation⁹² that lead to the deduction that situations can be created under which the action of enzymes mostly can be preserved in liquid/organic mixes at the temperature below 0 °C.

In the study, obvious advantage is the use of liquid media, of chloroplast reactions/response in evading the variations which is connected through subzero & permitting. The chloroplast diffusion, involves the small-scale molecules. The opportunity of perusing chloroplast’ reactions in a medium by a point at an icy temperature that is below H_2O is therefore possibly of excessive attention⁹³. This has been most disturbed the process of chloroplasts in solid (freezing) latency or very viscous resolutions, so they get the low temperatures glycerol’s. Newly stated light induced absorption variations at -40 °Celsius in chloroplasts deferent in 50 ~ volume per volume, of hygroscopic liquid⁹⁴. The solution of viscosity contains, ethylene glycol which is much, small that can cover glycerol. It has described that the photo-reduction of non-natural e^- accepting, which deferred the chloroplasts in liquid or organic combinations at subzero temperatures.

Conclusion:

Our surrounding has so much to impart us with so many things, and it owns not only advance structures but also effective useful units. These proceed decidedly enhanced the efficacy approaches in genetics to learn the functions of Chloroplast. While the succession of complete genome and the formation of high throughput apparatuses which examine the appearance, chloroplast study arrived at the genomic epoch. The green, Chloroplasts distinct arrangement of membrane bound organelles plastid, are the positions of photosynthesis and extra vigorous greenery purposes. Also, utter that the ‘Artificial Chloroplast’. for Light Driven Adenosine triphosphate (ATP) Synthesis. The learning concept of creation, would be broadly prolonged, and function and structure might be skilfully combined to figure out the orderly solar bio-chemical energy Alteration plans, which can benefit to boost the use of

insolation and environmental problems and ease the power. Besides that, the photo-reduction of non-natural e^- acceptors through, chloroplasts deferred in liquid or non- chemical combinations at subzero conditions.

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REFERENCE

- (1) Abdallah F: A prediction of the size and evolutionary origin of the proteome of chloroplasts of *Arabidopsis*. Trends Plant Sci. 5, 141–142 (2000).
- (2) The *Arabidopsis* Genome Initiative, Analysis of the genome sequence of the flowering plant *Arabidopsis thaliana*. Nature. 408, 796–815 (2000).
- (3) Rujan Tand Martin W: How many genes in *Arabidopsis* come from cyanobacteria? An estimate from 386 protein phylogenies. Trends Genet. 17, 113–120 (2001).
- (4) Martin W: Evolutionary analysis of *Arabidopsis*, cyanobacterial, and chloroplast genomes reveals plastid phylogeny and thousands of cyanobacterial genes in the nucleus. Proc. Natl Acad. Sci. USA 99, 12246–12251 (2002).
- (5) Higgs and DC: Inversions in the *Chlamydomonas* chloroplast genome suppress a petD 50 untranslated region deletion by creating functional chimeric mRNAs. Plant J. 14, 663–671 (1998).
- (6) Maenpaap: The ycf9 (orf 62) gene in the plant chloroplast genome encodes a hydrophobic protein of stromal thylakoid membranes. J. Exp. Bot. 51, 375–382 (2000).
- (7) RufS: A small chloroplast-encoded protein as a novel architectural component of the light-harvesting antenna. J. Cell Biol. 149, 369–378 (2000).
- (8) Baena-Gonzalez: Abnormal regulation of photosynthetic electron transport in a chloroplast ycf9 inactivation mutant. J. Biol. Chem. 276, 20795–20802 (2001).
- (9) Swiatek M: The chloroplast gene ycf9 encodes a photosystem II (PSII) core subunit, PsbZ, that participates in PSII supramolecular architecture. Plant Cell 13, 1347–1367 (2001).
- (10) Bennoun P and Delepelaire P: Isolation of photosynthetic mutants in *Chlamydomonas*. In Methods in Chloroplast Molecular Biology (Edelman, M., ed.), pp. 25–38, Elsevier (1982).
- (11) Miles: The role of high chlorophyll fluorescence photosynthesis mutants in the analysis of chloroplast thylakoid membrane assembly and function. Maydica 39, 35–45 (1994).
- (12) Meurer: Isolation of high-chlorophyll-fluorescence mutants of *Arabidopsis thaliana* and their characterisation by spectroscopy, immunoblotting and northern hybridisation. Planta 198, 385–396 (1996).
- (13) Meurer: A nuclear-encoded protein of prokaryotic origin is essential for the stability of photosystem II in *Arabidopsis thaliana*. EMBO J. 17, 5286–5297 (1998).

- (14) Lennartz: HCF164 encodes a thioredoxin-like protein involved in the biogenesis of the cytochrome b6f complex in *Arabidopsis*. Plant Cell 13, 2539–2551 (2001).
- (15) Kroll: VIPP1, a nuclear gene of *Arabidopsis thaliana* essential for thylakoid membrane formation. Proc. Natl Acad. Sci. USA 98, 4238–4242 (2001).
- (16) Settles: Sec-independent protein translocation by the maize Hcf106 protein. Science 278, 1467–1470 (1997).
- (17) Fisk DG: Molecular cloning of the maize gene *crp1* reveals similarity between regulators of mitochondrial and chloroplast gene expression. EMBO J. 18, 2621–2630 (1999).
- (18) Felder: The nucleus-encoded HCF107 gene of *Arabidopsis* provides a link between intercistronic RNA processing and the accumulation of translation-competent *psbH* transcripts in chloroplasts. Plant Cell 13, 2127–2141 (2001).
- (19) Schultes: Maize high chlorophyll fluorescent 60 mutation is caused by an *Ac* disruption of the gene encoding the chloroplast ribosomal small subunit protein 17. Plant J. 21, 317–327 (2000).
- (20) Niyogi: Chlamydomonas xanthophyll cycle mutants identified by video imaging of chlorophyll fluorescence quenching. Plant Cell 9, 1369–1380 (1997).
- (21) Niyogi and KK: *Arabidopsis* mutants define a central role for the xanthophyll cycle in the regulation of photosynthetic energy conversion. Plant Cell 10, 1121–1134 (1998).
- (22) Shikanai T: Identification and characterization of *Arabidopsis* mutants with reduced quenching of chlorophyll fluorescence. Plant Cell Physiol. 40, 1134–1142 (1999).
- (23) Li and XP: A pigment-binding protein essential for regulation of photosynthetic light harvesting. Nature 403, 391–395 (2000).
- (24) Munekage Y: Cytochrome b6f mutation specifically affects thermal dissipation of absorbed light energy in *Arabidopsis*. Plant J. 28, 351–359 (2001).
- (25) Varotto C: Identification of photosynthetic mutants of *Arabidopsis* by automatic screening for altered effective quantum yield of photosystem 2. Photosynthetica 38, 497–504 (2000).
- (26) Varotto C: Disruption of the *Arabidopsis* photosystem I gene *psaE1* affects photosynthesis and impairs growth. Plant J. 22, 115–124 (2000).
- (27) Pesaresi: Knock-out of the plastid ribosomal protein L11 in *Arabidopsis*: effects on mRNA translation and photosynthesis. Plant J. 27, 179–189 (2001).
- (28) Varotto C: The metal ion transporter IRT1 is necessary for iron homeostasis and efficient photosynthesis in *Arabidopsis thaliana*. Plant J. 31, 589–599 (2002).
- (29) Elrad: A major light-harvesting polypeptide of photosystem II functions in thermal dissipation. Plant Cell 14, 1801–1816 (2002).
- (30) N M Sadali, R G Sowden, Q Ling and R P Jarvis: Differentiation of chromoplasts and other plastids in plants. Plant Cell Rep. 38, 803–818 (2019).
- (31) P Jarvis and E López-Juez: Biogenesis and homeostasis of chloroplasts and other plastids. Nat. Rev. Mol. Cell Biol. 14, 787–802 (2013).
- (32) I. Egea: Chromoplast differentiation, Current status and perspectives. Plant Cell Physiol. 51, 1601–1611 (2010).
- (33) M Rodriguez: Concepcion A global perspective on carotenoids, Metabolism, biotechnology, and benefits for nutrition and health. Prog. Lipid Res. 70, 62–93 (2018).
- (34) T Sun: Carotenoid metabolism in plants, The role of plastids. Mol. Plant 11, 58–74 (2018).

- (35) G Giuliano: Provitamin A biofortification of crop plants. A gold rush with manyminers. *Curr. Opin. Biotechnol.* 44, 169–180 (2017).
- (36) B Llorente, JF Martinez-Garcia, C Stange and M. Rodriguez: Concepcion, Illuminatingcolors, Regulation of carotenoid biosynthesis and accumulation by light. *Curr. Opin.Plant Biol.* 37, 49–55 (2017).
- (37) J Lado, L Zacarías and MJRodrigo: Regulation of carotenoid biosynthesis during fruitdevelopment. *Subcell. Biochem.* 79, 161–198 (2016).
- (38) E T Wurtzel: Changing form and function through carotenoids and synthetic biology. *Plant Physiol.* 179, 830–843 (2019).
- (39) Boyer and PD: The ATP Synthase-a Splendid Molecular Machine.*Annu. Rev. Biochem.*66, 717–749 (1997).
- (40) Stock, D Leslie, A G W Walker and J E: Molecular Architecture ofthe Rotary Motor in ATP Synthase. *Science.* 286, 1700–1705 (1999).
- (41) Uchihashi, T Iino, R Ando and T Noji: High-Speed AtomicForce Microscopy Reveals Rotary Catalysis of Rotorless F1-ATPase.*Science.* 333, 755–758 (2011).
- (42) Richard, P Rigaud, J and L Graber: Reconstitution of CF^oF1 intoLiposomes Using a New Reconstitution Procedure. *Eur. J. Biochem.* 193, 921–925 (1990).
- (43) Luo, TJ M Soong, R Lan, E Dunn and BMontemagno: Photo-Induced Proton Gradients and ATP Biosynthesis Produced byVesicles Encapsulated in a Silica Matrix. *Nat. Mater.* 4, 220–224 (2005).
- (44) Choi andHJMontemagno: Artificial Organelle: ATPSynthesis from Cellular Mimetic Polymersomes. *Nano Lett.* 5,2538–2542 (2005).
- (45) Duan, LHe, Q Wang, K W Yan, X H Cui, YMöhwald and HLi: Adenosine Triphosphate Biosynthesis Catalyzed by FoF1 ATPSynthase Assembled in Polymer Microcapsules. *Angew. Chem.*119, 7126–7130. (2007).
- (46) Qi, W Duan, LWang, K W Yan, X H Cui, Y He and Q Li: Motor Protein CFoF1 Reconstituted in Lipid-Coated HemoglobinMicrocapsules for ATP Synthesis. *Adv. Mater.* 20, 601–605 (2008).
- (47) Li, JH Wang, Y F Ha, W Liu, Y Ding, L S Li and B J Zhang: Cyclodextrin-Based Microcapsules as Bioreactors for ATP Biosynthesis. *Biomacromolecules.* 14, 2984–2988 (2013).
- (48) Turina, P Samoray and D. Graber: H⁺ +/ATP Ratio of ProtonTransport Coupled ATP Synthesis and Hydrolysis Catalysed by CFoF1-Liposomes. *EMBO J.* 22, 418–426 (2003).
- (49) Duan, L Qi, W Yan, X H Cui, Y He, Q Wang, K W Li and DX Li: Proton Gradients Produced by Glucose OxidaseMicrocapsules Containing Motor FoF1-ATPase for Continuous ATPBiosynthesis. *J. Phys. Chem.*113, 395–399 (2009).
- (50) Racker and E Stoecken: Reconstitution of Purple MembraneVesicles Catalyzing Light-Driven Proton Uptake and AdenosineTriphosphate Formation. *J. Biol. Chem.* 249, 662–663 (1974).
- (51) Wendell, D Todd and J Montemagno: Artificial Photosynthesisin Ranaspumin-2 Based Foam. *Nano Lett.*10, 3231–3236 (2010).
- (52) Pitard, B Richard, P Dunach and M Rigaud: ATP Synthesis bythe FoF1 ATP Synthase from Thermophilic Bacillus PS3 Reconstitutedinto Liposomes with Bacteriorhodopsin. 2. Relationships betweenProton Motive Force and ATP Synthesis. *Eur. J. Biochem.* 235,779–788 (1996).
- (53) Feniouk, BA Cherepanov, D A Junge and W Mulkidjanian: Coupling of Proton Flow to ATP Synthesis in Rhodobacter Capsulatus:FoF1-ATP Synthase Is Absent from About Half of Chromatophores.*Biochim. Biophys. Acta, Bioenerg.*1506, 189–203 (2001).

- (54) Steinberg, Y G Rigaud, J L Durantini, E N Moore, A L Gust and D Moore: Light-Driven Production of ATP Catalysed by FoF₁-ATP Synthase in an Artificial Photosynthetic Membrane. *Nature* 392, 479–482 (1998).
- (55) Umena, Y Kawakami, K Shen, J R Kamiya: Crystal Structure of Oxygen-Evolving Photosystem II at a Resolution of 1.9 Å. *Nature* 473, 55–60 (2011).
- (56) Zhang, C X Chen, C H Dong, H X Shen, J R Dau, H Zhao and J Q: A Synthetic Mn₄Ca-cluster Mimicking the Oxygen-Evolving Center of Photosynthesis. *Science* 348, 690–693 (2015).
- (57) Daum, B Nicastro, D Il, J A McIntosh and J R Kuhlbrandt: Arrangement of Photosystem II and ATP Synthase in Chloroplast Membranes of Spinach and Pea. *Plant Cell*. 22, 1299–1312 (2010).
- (58) Nelson and N Yocum: Structure and Function of Photosystems I and II. *Annu. Rev. Plant Biol.* 57, 521–565 (2006).
- (59) Wang, W Y Chen, J Li, C Tian and W M: Achieving Solar Overall Water Splitting with Hybrid Photosystems of Photosystem II and Artificial Photocatalysts. *Nat. Commun.* 5, 8 (2014).
- (60) Yehezkeili, O Vered, R Tel Wasserman, J Trifonov, A Michaeli, D Nechushtai and R Willner: Integrated Photosystem II Based Photo-Bioelectrochemical Cells. *Nat. Commun.* 2012, 3, 742 (2012).
- (61) Andreiadis, E S Chavarot Kerlidou, M Fontecave and M Artero: Artificial Photosynthesis: from Molecular Catalysts for Light-driven Water Splitting to Photoelectrochemical Cells. *Photochem. Photobiol.* 87, 946–964 (2011).
- (62) Badura, A Kothe, T Schuhmann and W Rögner, M: Wiring Photosynthetic Enzymes to Electrodes. *Energy Environ. Sci.* 4, 3263–3274 (2011).
- (63) Kato and M Reisner: Protein Film Photoelectrochemistry of the Water Oxidation Enzyme Photosystem II. *Chem. Soc. Rev.* 43, 6485–6497 (2014).
- (64) Kopnov, F Cohen Ofri and I Noy: Electron Transport between Photosystem II and Photosystem I encapsulated in Sol-Gel Glasses. *Angew. Chem., Int. Ed.* 50, 12347–12350 (2011).
- (65) Carpentier R: *Photosynthesis Research Protocols*; Humana Press Inc. Totowa (2011).
- (66) Vullev VI: From biomimesis to bioinspiration: what's the benefit for solar energy conversion application? *J Phys Chem Lett* 2:503e8 (2011).
- (67) Nealon KH Conrad PG: Life-past, present and future. *Philos Trans R Soc Lond B Biol Sci.* 354:1923e39 (1999).
- (68)a) Anderson JM: Photoregulation of the composition, function, and structure of thylakoid membranes. *Ann Rev Plant Physiol.* 37:93e136 (1986).
- B) Ruban AV, Johnson MP, Duffy CDP: Natural light harvesting: principles and environmental trends. *Energy Environ Sci.* 4:1643e50 (2011).
- (69) Gust D, Kramer D, Moore A, Moore TA and Vermaas W: Engineered and artificial photosynthesis: human ingenuity enters the game. *MRS Bull.* 33:383e7 (2008).
- (70) Manga KK, Zhou Y, Yan YL, Loh KP: Multilayer hybrid films consisting of alternating graphene and Titania nanosheets with ultrafast electron transfer and photoconversion properties. *Adv Funct Mater.* 19:3638e43 (2009).
- (71)a) Lomoth R, Magnuson A, Sjödin M, Huang P, Styring S and Hammarström L: Mimicking the electron donor side of photosystem II in artificial photosynthesis. *Photosynth Res.* 87:25e40 (2006).
- b) Hammarström L and Hammes-Schiffer S: Artificial photosynthesis and solar fuels. *Acc Chem Res.* 42:1859e60 (2009).

- (72) Meunier CF, Rooke JC, Le´onard A, Xie H and Su BL: Living hybridmaterials capable of energy conversion and CO₂assimilation. *Chem Commun* .46:3843e59 (2010).
- (73) Modesto-Lopez LB, Thimsen EJ, Collins AM, Blankenship RE and Biswas P: Electrospray-assisted characterization anddeposition of chlorosomes to fabricate a biomimetic lightharvesting device. *Energy Environ Sci*. 3:216e22 (2010).
- (74) a)Liu CJ, Burghaus U, Besenbacher F and Wang ZL:Preparationand characterization of nanomaterials for sustainableenergy production. *ACS Nano*. 4:5517e26 (2010).
- b) Kim JH, Lee M, Lee JS and Park CB: Self-assembled lightharvesting peptide nanotubes for mimicking naturalphotosynthesis. *Angew Chem*. 124:532e5 (2012).
- (75) Bard AJ and Fox MA: Artificial photosynthesis: solar splitting ofwater to hydrogen and oxygen. *Acc Chem Res*. 28:141e5 (1995).
- (76) Wu YM, Xing MY, Zhang JL and Chen F: Effective visible lightactive boron and carbon modified TiO₂ photocatalyst for degradation of organic pollutant. *Appl Catal B Environ*. 97:182e9 (2010).
- (77) Fujishima A and Honda K: Electrochemical photolysis of water at a semiconductor electrode. *Nature*. 238:37e8 (1972).
- (78) Song MY, Kim DK, Ihn KJ, Jo SM and Kim DY: Electrospun TiO₂electrodes for dye-sensitized solar cells. *Nanotechnology*. 15:1861e5 (2004).
- (79) Song YY, Schmidt-Stein F, Berger S, Schmuki P: TiO₂ nano test tubes as a self-cleaning platform for high-sensitivity immunoassays. *Small*. 6:1180e4 (2010).
- (80) Ihara T, Miyoshi M, Iriyama Y, Matsumoto O and Sugihara S: Visible-light-active titanium oxide photocatalyst realized byan oxygen-deficient structure and by nitrogen doping. *ApplCatal Be Environ* 42:403e9 (2003).
- (81) Choi WY, Termin A and Hoffmann MR: The role of metal iondopants in quantum sized TiO₂: correlation betweenphotoreactivity and charge carrier recombination dynamics.*J Phys Chem*. 98:13669e79 (1994).
- (82) Huang BS, Chang FY and Wey MY: Photocatalytic properties ofredox-treated Pt/TiO₂photocatalysts for H₂ production froman aqueous methanol solution. *Int J Hydrogen Energy*. 35:7699e705 (2010).
- (83) Li XF, Fan TX, Zhou H, Chow SK, Zhang W, Zhang D: light-harvesting and photocatalytic properties inmorph-TiO₂ from green-leaf biotemplates. *Adv Funct Mater*. 19:45e56 (2009).
- (84) Mor GK, Varghese OK, Paulose M, Shankar K and Grimes CA: Areview on highly ordered, vertically oriented TiO₂ nanotubearrays: fabrication, materials properties, and solar energyapplications. *Sol Energy Mater Sol Cells*. 90:2011e75 (2006).
- (85) Feng XJ, Shankar K, Varghese OK, Paulose M, Latempa TJ andGrimes CA: Vertically aligned single crystal TiO₂ nanowire arrays grown directly on transparentConducting oxide coated glass: synthesis details and applications. *Nano lett*. 3781-6 (2008).
- (86) Lakshmi BB, Patrissi CJ and Martin CR: Solgel templatesynthesis of semiconductor oxide micro-andnaonostructures. *Chem Mater*. 9:2544e50 (1997).
- (87) Shi N, Li XH, Fan TX, Zhou H, Ding J and Zhang D: BiogenicN-I-codoped TiO₂ photocatalyst derived from kelp forefficient dye degradation. *Energy Environ Sci*. 4:172e80 (2011).
- (88) Zhang W, Zhang D, Fan TX, Gu J, Ding J and Wang H: Novelphotoanode structure templated from butterfly wing scales.*Chem Mater*. 21:33e40 (2009).
- (89) a)Zscheile FP and Comar CL: Influence of preparative procedureon the purity of chlorophyll components as shown byabsorption spectra. *Bot Gaz*. 102:463e81 (1941).

b) Croce R, Cinque G, Holzwarth AR and Bassi R: The soresorption properties of carotenoids and chlorophylls in antenna complexes of higher plants. *Photosynth Res.* 64:221e31 (2000).

(90) Dutton PL, Leigh JS and Seibert M: Primary processes in photosynthesis: in situ ESR studies on the light induced oxidized and triplet state of reaction center bacteriochlorophyll. *Biochem Biophys Res Commun.* 46:406e13 (1972).

(91) Hui Bon, Hoa G and Douzou, J: *Biol. Chem.* 248, 4649-4654 (1973).

(92) Douzou P: *Mol. Cell. Biochem.* 1, 15-27 (1973).

(93) Inou H, and Nishimura M: *Plant. Cell. Physiol.* 12, 137-145 (1971).

(94) Ames, J., Pulles, M. P. J. and Velthuis BR: *Biochim. Biophys. Acta* 325, 472-482 (1973).