

# Hepatoprotective and Antioxidant Activity of Edible Mushroom *Termitomyces Hemii* in CCl<sub>4</sub>-Induced Hepatotoxicity in Albino Rats

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## Abstract:

The present study included the assessment of hepatoprotective efficacy of the edible mushroom *Termitomyces hemii* against Carbon Tetrachloride (CCl<sub>4</sub>) hepatotoxicity in albino Wistar rats. The aqueous extract of the fruiting bodies showed the presence of several pharmacologically important biochemical constituent compounds including flavonoids, alkaloids, phenolics, tannins, saponins, among others. The in-vitro antioxidant analysis of the extract showed a strong free radical scavenging activity against DPPH (1, 1- diphenyl-2- picryl hydrazyl) radicals using Ascorbic acid as reference standard. At 100 µg/ml concentration, the extract exhibited 47.58% DPPH radical scavenging activity in comparison to the 55.14% DPPH radical scavenging activity of the standard reference Ascorbic acid. The in-vivo studies showed that a dose of 500 mg/Kg Body weight/day of the extract in CCl<sub>4</sub>-induced hepatotoxic rats resulted into significant ( $p \leq 0.05$ ) improvement in the blood levels of liver function parameters, such as bilirubin, serum albumin, total protein, aspartate aminotransferase (AST), alanine transaminase (ALT), and alkaline phosphatase (ALP). The marked hepatoprotective efficacy of the present mushroom may be attributed to its rich content of bioactive and potent biochemical constituent compounds having marked antioxidant potentialities.

**Key words:** Mushroom, Hepatoprotective, Antioxidant, *Termitomyces hemi*, Carbon tetrachloride

## Introduction:

Mushrooms are actually the macrofungi with fruiting bodies large enough to be seen through naked eyes, which can be either epigeous or hypogeous. They belong to two major groups, Ascomycota and Basidiomycota, and possess variable degree of edibility. Studies have reported that there are around 140,000 species of macrofungi found across the globe, out of which only 10% (14,000) species have been described to date (Chang and Miles, 2004). Among these, approximately 2000 species are considered as edible, and about 700 species have been considered to possess medicinal or pharmacological properties (Chang and Miles, 2004; Karaman *et al.*, 2012). The edible mushrooms have been reported to contain essential mineral nutrients, fibres, amino acids, fatty acids, vitamins, polysaccharides etc., and are considered to be

a rich source of proteins and bioactive secondary metabolites like alkaloids, flavonoids, terpenoids, tannins, saponins, among others (Kues and Liu, 2000; Mattila *et al.*, 2002; Aidaa *et al.*, 2009). Mushrooms are typically regarded as functional foods or nutraceutical products since they are nutritionally beneficial when consumed regularly or when their isolated bioactive constituent compounds are consumed (Lakhanpal and Rana, 2005).

The liver is a complex organ of the body, which is involved in carrying out a number of vital metabolic and physiological functions, such as drug elimination and detoxification, metabolism of food molecules and maintenance of blood glucose levels by glycogenesis and glycogenolysis, synthesis and secretion of bile, synthesis of several plasma proteins including blood clotting factors and many others physiological and metabolic functions (Gowri Shankar *et al.*, 2008). Liver damage or liver related disorders, therefore may produce serious health consequences, sometimes may prove fatal. Liver damage is a prevalent and widespread condition that typically entails oxidative stress and is marked by a steady progression from steatosis to chronic hepatitis, fibrosis, cirrhosis, and hepatocellular cancer (Kodavanti *et al.*, 1989). Carbon Tetrachloride (CCl<sub>4</sub>), a toxic substance to the liver, is commonly employed in scientific research to induce hepatotoxicity in animal models and to assess the efficacy of any hepatoprotective agent or medicines (Seifert, 1994). The induction of hepatotoxicity by CCl<sub>4</sub> includes the binding with Cytochrome P450 and thereby release of highly reactive free radical i.e. Trichloromethyl (CCl<sub>3</sub>·), which then mediates cascade of reactions leading to enhanced lipid peroxidation and cellular damage, consequently leading to hepatocellular injury and hepatic necrosis (Kadhwa *et al.*, 2000). Any chronic or acute hepatotoxicity produced by the hepatotoxic agent leads to the abnormal levels of specific biochemical parameters or enzymes in the blood or liver tissues, such as bilirubin, serum albumin, total protein, aspartate aminotransferase (AST), alanine transaminase (ALT), and alkaline phosphatase (ALP) (Giannini *et al.*, 2005; Maity and Ahmad, 2012).

*Termitomyces hemii* is an edible macrofungus belonging to family Lyophyllaceae, and is commonly found in South Asia. This fungus lives in a symbiotic association in or on termite nests. Puttaraju *et al.*, (2006) studied the phenolic content and antioxidant efficacy of 23 species of edible mushrooms collected from different locations of India, and found *T. hemi* as the best variety with highest phenolic content and most prominent free radical scavenging activity. The marked antioxidant efficacy of *T. hemi* was attributed to its rich phenolic content. Since mushrooms generally contain a number of potent biochemical components with several pharmacologically important functions, the present work had been undertaken to qualitatively analyze the mycochemical constituent compounds and to determine the antioxidant potentiality and hepatoprotective efficacy of the present edible mushroom *T. hemi*.

## **Materials and Methods:**

### **Collection of sample and preparation of extract:**

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Fresh fruiting bodies of *T. hemi* have been collected from East Singhbhum region of Jharkhand, India and brought to the laboratory. The fruiting bodies were thoroughly washed, disinfected with 0.1% HgCl<sub>2</sub> followed by repeated washing with deionized water. Now the fruiting bodies were dried under shade for 7-8 days, and upon complete drying, and then powdered using an electric grinder. The powder was then sieved and stored for further use. 20 gm of the powdered sample was subjected to solvent extraction using water (500 ml) as solvent with the help of Soxhlet apparatus. The extract was then filtered and stored for further use.

### **Qualitative biochemical analysis of *T. hemi* extract:**

The preliminary qualitative analysis for the presence of different secondary metabolites was done according to previously established standard tests (Bhaskar and Kumar, 2012).

### **Assessment of Antioxidant activity:**

The antioxidant activity of the extract was determined by DPPH (1, 1- diphenyl-2- picryl hydrazyl) radical scavenging assay following the previously established standard methods (Moon and Terao, 1998; Kumar et al., 2008). Different quantities of samples (10µg, 50µg, and 100µg) were taken in Dimethyl Sulfoxide (DMSO) and methanol was added to bring the volume up to 500µl. These tubes were then filled with 5 ml of a 0.1 mM methanolic solution of 1,1-diphenyl-2-picryl hydrazyl (DPPH; Sigma-Aldrich, Bangalore), followed by vigorous shaking. A control solution was kept with the equal amount of methanol but without the test component. The tubes were let to stand at room temperature for 20 minutes. At 517 nm, the samples' absorbance was measured. The reference standard used was butylated hydroxy anisole (BHA).

Free Radical scavenging activity was calculated using the following formula:

$$\% \text{ radical scavenging activity} = \frac{(\text{control OD} - \text{sample OD})}{\text{control OD}} \times 100.$$

### **Animals and acute toxicity studies:**

Adult albino wistar rats (*Rattus norvegicus*) weighing between 180-220 gm were maintained at a temperature of 25±5°C and a relative humidity of 50±15% over a paddy husk bed under standard laboratory conditions. The rats were fed with commercial pellet diet and water ad libitum. The experiment was undertaken with prior approval from “Institutional Ethical Committee” of Ranchi University, Jharkhand, India. The staircase method was used for acute toxicity studies following the OECD guidelines (2004). The rats were divided into five groups of ten rats each and fed with increasing concentration of the extract. No mortality was observed upto the concentration of 2000mg/kg Body Weight per day within a period of 48 hrs.

### **Animal groups and Research design:**

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A mixture of CCl<sub>4</sub> (30%) and liquid paraffin (1:2 V/V) was injected intra-peritoneally (i.p.) every 72 hrs. for 14 days. The animals were divided into three groups with ten rats in each group and the experiment was carried out as follows:

Group A (Control): received 1 ml of normal saline orally

Group B (CCl<sub>4</sub> treated): received 1ml/Kg i.p. of CCl<sub>4</sub> every 72 hrs

Group C (CCl<sub>4</sub> treated + Extract): hepatotoxic rats, received 500 mg/BW/day of *T. hemii* extract

The experiment was continued for 14 days. At the end of 14<sup>th</sup> day, all experimental animals were kept fasting overnight, and then the blood samples were collected randomly from each group in triplicates. The blood samples were centrifuged at 3000 rpm for 10 minutes and the clear serum was collected for further estimation of biochemical parameters and enzymes. Serum ALT (U/L) and AST (U/L) were estimated using the Reitman and Frankel (1957) method, total protein and albumin were assessed using the Kingsley and Frankel (1939) method, and serum AST (U/L) was quantified using the method of Bessey *et al.* (1964).

## Results:

### Qualitative biochemical analysis of *T. hemi* extract:

The result of qualitative analysis of secondary metabolite content of the aqueous extract of fruiting bodies of *T. hemi* has been shown in Table 1.

Table 1: Showing qualitative analysis of aqueous extract of *T. hemii* extract

Biochemical constituents	Presence/Absence (+/-)
Reducing sugars (Carbohydrates)	+
Proteins	+
Amino acids	+
Alkaloids	+
Flavonoids	+
Phenolics	+
Saponins	+
Tannins	+
Sterols and steroids	+
Quinons	+

### Antioxidant activity of *T. hemi* extract:

Table 2 shows the DPPH radical scavenging activity of the *T. hemi* extract using Ascorbic acid as the standard reference.

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Table 2: Showing % DPPH radical scavenging activity at various concentrations of *T. hemi extract*

Concentration ( $\mu\text{g/ml}$ )	% Free radical scavenging activity of Pt-AgNPs	% Free radical scavenging activity of Ascorbic acid
25	24.57	31.63
50	37.91	44.21
100	47.58	55.14

**Hepatoprotective activity of *T. hemi* extract:**

The results of analysis of hepatoprotective activity of *T. hemi* extract have been shown in Table 3.

Table 3: Showing hepatoprotective effect of *T. hemii* extract on serum levels of liver function parameters in CCl<sub>4</sub>-induced hepatotoxic rats

Animal Groups	AST (IU/L)	ALT (IU/L)	ALP (IU/L)	Total Protein (g/dL)	Albumin (g/dL)	Total Bilirubin (mg/dL)
Group 1	63.34 $\pm$ 7.16	56.32 $\pm$ 6.71	127.49 $\pm$ 9.53	21.97 $\pm$ 3.29	7.08 $\pm$ 0.64	1.06 $\pm$ 0.18
Group 2	183.29 $\pm$ 9.87 <sup>a</sup>	189.73 $\pm$ 12.39 <sup>a</sup>	214.37 $\pm$ 13.89 <sup>a</sup>	6.58 $\pm$ 0.72 <sup>a</sup>	3.04 $\pm$ 0.52 <sup>a</sup>	3.18 $\pm$ 0.49 <sup>a</sup>
Group 3	71.59 $\pm$ 8.33 <sup>b</sup>	72.19 $\pm$ 8.37 <sup>ab</sup>	131.79 $\pm$ 10.21 <sup>b</sup>	17.39 $\pm$ 3.11 <sup>ab</sup>	6.22 $\pm$ 0.58 <sup>b</sup>	1.51 $\pm$ 0.23 <sup>ab</sup>
Group 4	69.11 $\pm$ 8.59 <sup>b</sup>	69.27 $\pm$ 7.13 <sup>ab</sup>	125.63 $\pm$ 11.59 <sup>b</sup>	20.46 $\pm$ 2.58 <sup>b</sup>	7.37 $\pm$ 0.79 <sup>b</sup>	1.26 $\pm$ 0.36 <sup>b</sup>

\*a (significantly different from Group 1); b (significantly different from Group 2) at  $p \leq 0.05$

**Discussion:**

The administration of CCl<sub>4</sub> results into abnormal rise in oxidative stress within the hepatic cells or tissues, which leads to enhanced lipid peroxidation and thereby damage to the hepatocellular membranes (De Groot et al., 1988; Acharya *et al.*, 2012). The degeneration of hepatocellular membranes and the obstructions in the biliary system of the liver results into an abnormal increase in the blood levels of AST, ALT, ALP and bilirubin (Huo *et al.*, 2011). The oxidative stress can also produce damage in the intracellular structures like endoplasmic reticulum, mitochondria along with DNA within the hepatic cells, and consequently the reduced protein synthesis, which results into abnormal decrease in the blood levels of proteins and albumin (Rajendran *et al.*, 2009). In the present work the administration of *T. hemi* extract in hepatotoxic group of rats had resulted into significant ( $p \leq 0.05$ ) decrement in the blood levels of AST, ALT, ALP, and bilirubin, and a concurrent and significant ( $p \leq 0.05$ ) rise in the blood levels of albumin and proteins, back towards their normal values, indicating the protective effect of *T. hemii* extract against the CCl<sub>4</sub>-induced hepatotoxicity. Depletion in the enhanced levels of bilirubin along with decrease in the blood levels of ALP indicates the efficacy of *T. hemi* extract

to stabilize biliary disfunctions. Further, the depletion in the blood levels of AST, ALT and bilirubin suggests that the *T. hemii* extract has the ability to minimize the structural damages in the hepatocytes, as well as to accelerate the regeneration in hepatic cells and tissues.

Several previous works have reported that the macrofungi or mushrooms are rich in their potent and bioactive secondary metabolite content, which have marked pharmacological properties (Nada *et al.*, 2010). Previous studies reported that the mushrooms extracts or specific biochemical constituents isolated from mushroom extract showed significant hepatoprotective properties. The edible mushrooms like *Lentinus edodes*, *Grifola frondosa*, *Tricholoma lobayense*, *Ramaria botrytis*, *Calocybe indica* and *Astraeus hygrometricus* are reported to possess marked Hepatoprotective efficacies (Ooi Vec, 1996; Kim *et al.*, 2003; Chatterjee *et al.*, 2011). Ajith *et al.*, (2006) had reported the Hepatoprotective efficacy of the edible mushroom *Phellinus rimosus* against CCl<sub>4</sub>-mediated hepatotoxicity in rats. Jayakumar *et al.*, (2006) had reported that the extract of edible mushroom *Pleurotus ostreatus* shows marked improvement in the hepatotoxicity caused by CCl<sub>4</sub>. Acharya *et al.*, (2012) had reported the Hepatoprotective efficacy of the edible mushroom *Macrocybe gigantean* against CCl<sub>4</sub>-induced hepatotoxicity in rats. Other works have also reported that edible mushrooms like *Agaricus blazei* (Al-Dbass *et al.*, 2012), *Russula albonigra* (Chaterjee *et al.*, 2012), and *Pleurotus cornucopiae* (El-Bohi *et al.*, 2009), possess the ability to improve the liver damage from CCl<sub>4</sub>-induced hepatotoxicity in rats.

The studies have reported that the marked hepatoprotective efficacy of mushroom extracts is primarily due to their strong antioxidant potentiality, which can be attributed to their bioactive chemical constituent compounds such as flavonoids, phenolics, alkaloids, among others (Di Carlo *et al.*, 1999; Al-Dbass *et al.*, 2012; Acharya *et al.*, 2012). The qualitative analysis of the present mushroom *Termitomyces hemii* had shown the presence of potent antioxidant compounds like flavonoids, alkaloids, phenolics etc. in the extract. Further, the results of the present study also showed the strong antioxidant activity of *T. hemii* extract. Hence, the marked hepatoprotective efficacy of the present edible mushroom, i.e., *T. hemii*, can be attributed to its bioactive chemical constituents or the secondary metabolite content.

### **Conclusion:**

From the results of the present work, it can be concluded that the aqueous extract of *T. hemii* fruiting bodies has marked hepatoprotective activity against CCl<sub>4</sub>-induced hepatotoxicity in the present mammalian animal model, i.e., albino Wistar rats, which might be due to its strong antioxidant properties. Hence, the present edible mushroom, i.e., *T. hemii*, can be a potent nutraceutical dietary source, which can further be studied and explored to isolate and develop new drugs and medicinal agents from this mushroom.

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## References:

1. Acharya, K., Chatterjee, S., Biswas, G., Chatterjee, A., & Saha, G. K. (2012). Hepatoprotective effect of a wild edible mushroom on carbon tetrachloride-induced hepatotoxicity in mice. *Int J Pharm Pharm Sci*, 4(3), 285-288.
2. Aidaa, F.M.N.A, Shuhaimia, . M. , Yazidb, M. and Maarufc, A.G. 2009. Mushroom as a potential source of prebiotics: a review. *Trends in Food Science & Technology* 20; 567-575. Elsevier Pubn.
3. Ajith TA, Sheena N, Janardhanan KK. *Phellinus rimosus* protects carbon tetrachloride-induced chronic hepatotoxicity in rats: antioxidant defense mechanism. *Pharm Biol* 2006; 44: 467–74.
4. Al-Dbass, A. M., Al-Daihan, S. K., & Bhat, R. S. (2012). *Agaricus blazei* Murill as an efficient hepatoprotective and antioxidant agent against CCl<sub>4</sub>-induced liver injury in rats. *Saudi journal of biological sciences*, 19(3), 303-309.
5. Bessey, O.A., Lowry, D.H., Brock, M.J. (1964). A method for the rapid determination of alkaline phosphatase with five cubic millimeter of serum. *J. Biol. Chem.* 164: 321-326
6. Bhaskar, A. and Kumar, A. 2012. Antihyperglycemic, antioxidant and hypolipidimic effect of *Punica granatum* L. flower extract in streptozotocin induced diabetic rats. *Asian Pacific J. Trop. Biomed.*, pp. s1764-s1769.
7. Chang, S.T. and Miles, P.G. (2004). Mushrooms cultivation, Nutritional value, Medicinal effect, and Environmental impact. *United states: CRC press*.
8. Chatterjee S, Dey A, Dutta R, Dey S, Acharya K. Hepatoprotective Effect of the Ethanolic Extract of *Calocybe indica* on Mice with CCl<sub>4</sub> Hepatic Intoxication. *Int J PharmTech, Res* 2011; 3: 2162–8.
9. Chatterjee, S., Datta, R., Dey, A., Pradhan, P., & Acharya, K. (2012). In vivo hepatoprotective activity of ethanolic extract of *Russula albonigra* against carbon tetrachloride-induced hepatotoxicity in mice. *Research Journal of Pharmacy and Technology*, 5(8), 1034-1038.
10. De Groot, H., Littauer, A., Hugo-Wissemann, D., Wissemann, P., Noll, T. (1988). Lipid peroxidation and cell viability in isolated hepatocytes in a redesigned oxystat system: Evaluation of the hypothesis that lipid peroxidation, preferentially induced at low oxygen partial pressure, is decisive for CCl<sub>4</sub> liver cell injury . *Arch Biochem Biophys.* 264:591-99.
11. Di Carlo G, Mascola N, Izzo AA, Capasso F. Flavonoids: old and new aspects of a class of natural therapeutic drugs. *Life Sci* 1999; 65: 337–53.
12. El Bohi, K. M., Hashimoto, Y., Muzandu, K., Ikenaka, Y., Ibrahim, Z. S., Kazusaka, A., ... & Ishizuka, M. (2009). Protective effect of *Pleurotus cornucopiae* mushroom extract on carbon tetrachloride-induced hepatotoxicity. *Jpn. J. Vet. Res*, 57(2), 109-118.

13. Giannini, E.G.; Testa, R.; Savarino, V. Liver enzyme alteration: A guide for clinicians. *Can. Med. Assoc. J.* 2005, *172*, 367–379
14. Gowri-Sankar, N.L.; Manavalan, R.; Venkappayya, D.; Raj, C.D. Hepatoprotective and antioxidant effects of *Commiphora berryi* (Arn) Engl bark extract against CCl<sub>4</sub>-induced oxidative damage in rats. *Food Chem. Toxicol.* 2008, *46*, 3182–3185.
15. Huo, H.Z., Wang, B., Liang, Y.K., Bao, Y.Y., Gu, Y. (2011). hepatoprotective and antioxidant effects of licorice extract against CCl<sub>4</sub>-induced oxidative damage in rats. *Int. J. Mol. Sci.* 12: 6529-6543
16. Jayakumar, T., Ramesh, E., & Geraldine, P. (2006). Antioxidant activity of the oyster mushroom, *Pleurotus ostreatus*, on CCl<sub>4</sub>-induced liver injury in rats. *Food and chemical toxicology*, *44*(12), 1989-1996.
17. Kadhaka, M.B.; Gladen, B.C.; Baird, D.D.; Dikalova, A.F.; Sohal, R.; Hatch, G.E.; Jones, D.P.; Mason, R.P.; Barreti, J.C. Biomarkers of oxidative stress study: Are plasma antioxidants markers of CCl<sub>4</sub> poisoning? *Free Radic. Biol. Med.* 2000, *28*, 838–845
18. Karaman, M., Vesic, M., Stahl, M., Novakovic, M., Janjic, L., Matavuly, M. (2012). Bioactive properties of wild growing mushroom species *Ganoderma applanatum* (Pers.) pat. From Fruska Gora forest (Serbia). *Ethnomed. Ther. Valid.* 32: 361-377.
19. Kingsley, S.R. and Frankel, S.J. (1939). The determination of serum total protein albumin and globulin by biuret reaction. *J. Biol. Chem.* 128:131-137.
20. Kim HJ, Lee KR. Effect of *Ramaria botrytis* methanol extract on antioxidant enzyme activities in benzo(a)pyrene-treated mice. *Korean J Food Sci Technol* 2003; *354*: 286–90.
21. Kues, U.; Liu, Y. Fruiting body production in basidiomycete. *Appl. Microbiol. Biotechnol.* 2000, *54*, 141–152.
22. Kodavanti, P.R.; Joshi, U.M.; Young, Y.A.; Meydrech, E.F.; Mehendale, H.M. Protection of hepatotoxic and lethal effects of CCl<sub>4</sub> by partial hepatectomy. *Toxicol. Pathol.* 1989, *17*, 494–505
23. Kumar S., Kumar D., Manjusha, Saroha K., Singh N. and Vashishta B. 2008. Antioxidant and free radical scavenging potential of *Citrullus colocynthis* (L.) Schrad. methanolic fruit extract, *Acta. Pharm.* 58; 215–220.
24. Lakhanpal, T.N.; Rana, M. Medicinal and nutraceutical genetic resources of mushrooms. *Plant Gen. Res.* 2005, *3*, 288–303.
25. Mattila, P.; Salo-Vaananen, P.; Konko, K.; Aro, H.; Jalava, T. Basic composition and amino acid contents of mushrooms cultivated in Finland. *J. Agric. Food Chem.* 2002, *50*,



- 6419–6422.Moon, J.H. and Tearo, J. (1998). Antioxidant activity of caffeic acid and dihydrocaffeic acid in lard and human low density protein. *J. Agri. And Food Chem.* 46: 5062-65.
26. Nada, S. A., Omara, E. A., Abdel-Salam, O. M., & Zahran, H. G. (2010). Mushroom insoluble polysaccharides prevent carbon tetrachloride-induced hepatotoxicity in rat. *Food and Chemical Toxicology*, 48(11), 3184-3188.
27. OECD (2004). OECD guidelines for the testing of chemicals /section 4: Health effects test no. 423; Acute Oral Toxicity-Acute Toxic Class Method.Organisation for Economic Cooperation and Development.
28. Ooi VEC. Hepatoprotective effect of some edible mushrooms. *Phytother Res* 1996; 10: 536–8.
29. Puttaraju, N. G., Venkateshaiah, S. U., Dharmesh, S. M., Urs, S. M. N., & Somasundaram, R. (2006). Antioxidant activity of indigenous edible mushrooms. *Journal of agricultural and food chemistry*, 54(26), 9764-9772.
30. Rajendran R, Hemalatha S, Akasakalai K, Madhukrishna CH, Vittal BS and Sundaram RM. Hepatoprotective activity of Mimosa pudica leaves against carbontetrachloride induced toxicity. *Journal of Natural Products* 2009; 2: 116-122.
31. Reitman, S. and Frankel, S.A. (1957). Colorimetric method for the determination of serum glutamic oxaloacetic and pyruvic transaminase. *Am. J. Clin. Path.* 28:56
32. Seifert, W.F., Bosma, A., Brouwer, A. (1994). Vitamin A deficiency potentiates CCl<sub>4</sub>-induced liver fibrosis in rats. *Hepatology*, 19(1): 193-201.
33. Wasser, S.P. and A. Weis, (1999). Medicinal properties of substances occurring in higher basidiomycetes mushrooms: current perspectives (review). *Int. J. Med. Mushrooms.*, 1:31-62.